APPENDIX A Glossary

A.1 Glossary of Terms and Acronyms

GLOSSARY OF TERMS AND ACRONYMS[†]

Asymptomatic: Showing no symptoms of illness.

Attack rate: The occurrence of disease observed among a defined population over a limited period of time.

Bacterium: A one-celled living microorganism that can cause foodborne infections and intoxications.

Bare hand contact: Having bare hands in direct contact with prepared or ready-to-eat food items.

Biological hazard*: A bacterial, viral, or parasitic agent that may make food unsafe to eat. Often associated with microorganisms naturally found in raw meat and poultry products or microorganisms introduced during processing of meat and poultry products.

Carrier: Individuals who harbor an infectious agent but are asymptomatic.

Case definition: A set of criteria for determining who should be classified as a case. The definition is comprised of clinical information and should include information related to time, place, and person.

Case: A person with the particular item of interest or disease.

Case-control study: An observational, analytical study in which individuals are not part of a well-defined population. Inclusion into the study is dependent upon the presence or absence of illness or disease.

Chemical hazard*: Chemicals that are naturally occurring in foods (i.e., aflatoxins, mucotoxins, and shellfish toxins) or added during the processing of foods. Intentional or unintentionally added chemicals may include components of animal feed or drinking water, animal drugs, pesticides, food ingredients themselves, or chemicals used in the processing establishment, like lubricants, cleaners, paints, and coatings.

Cohort study: An observational, analytical study involving a well-defined group of individuals. Inclusion into the study is dependent upon a common experience of exposure.

Common-source outbreak: A type of outbreak that occurs when individuals are exposed to some point-source of infection at the same time.

[†] Many definitions were taken from (1) Last, JM ed. *A Dictionary of Epidemiology*, 3rd ed. New York: Oxford U. Press, 1995 and (2) Chin, J ed. *Control of Communicable Diseases Manual*, 17th ed. Washington DC: American Public Health Association, 2000.

^{*} USDA – FSIS. Guidebook for the Preparation of HACCP Plans. September 1999.

Confidence intervals: A range of numbers that indicates how "confident" one can be that the observed number actually lies within the range.

Confidentiality: The obligation to not disclose identifying information unless needed to protect the public's health.

Confirmed disease outbreak: A foodborne disease outbreak in which laboratory analysis of appropriate specimens identifies a causative agent and epidemiologic analysis implicates the food as the source of the illness. [NOTE: Positive laboratory identification of the disease-causing organism is not necessary to determine that a foodborne disease outbreak has occurred nor is this identification needed to begin investigation.]

Consumer: A person who is a member of the public, takes possession of food, is not functioning in the capacity of an operator of a food establishment or food processing plant, and does not offer the food for resale.

Continual-source outbreak: A type of outbreak that occurs when a source remains contaminated and exposure and illness continues.

Control: In a case-control study, a person without illness or disease.

Control measure[‡]: Any action or activity that can be used to prevent, eliminate, or reduce a significant hazard.

Control point[‡]: Any step at which biological, chemical, or physical factors can be controlled.

Corrective action[‡]: Procedures that are initiated when a deviation or problem in the flow of food preparation is identified.

Critical limit[‡]: The maximum and/or minimum value at which a biological, chemical, or physical hazard must be controlled at a given critical control point to ensure food safety.

Critical control point (CCP)[‡]: A step at which control can be applied to prevent or eliminate a food safety hazard or reduce it to an acceptable level.

Cross-contamination: The transfer of pathogens from one food item to another food item during food preparation through cooking equipment, utensils, and the hands of food handlers.

Deviation[‡]: Failure to meet a critical limit.

Epidemic curve (epi curve): A histogram or graph that provides a visual depiction of the outbreak over time.

Epidemiology: The study of the distribution and determinants of health-related states or events within a specific population, and the application of this study to control health problems.

FAT TOM: Conditions that favor the growth of foodborne microorganisms within the environmental setting, specifically food, acidity, time, temperature, oxygen, and moisture.

Fecal-oral route: The ingestion of stool from an infected person or animal through food, liquids or direct contact.

Food establishment: An operation that stores, prepares, packages, serves, vends, or otherwise provides food for human consumption. Food establishments include a restaurant; satellite or catered feeding location; catering operation if the operation provides food directly to a consumer or to a conveyance used to transport people; market; vending location; conveyance used to transport people; institution; or food bank.

Food handler: A person who directly handles or prepares food.

Food processing plant: A commercial operation that manufactures, packages, labels, or stores food for human consumption and does not provide food directly to a consumer.

Foodborne disease outbreak (FBDO): (1) Two or more individuals (from different households) who experience a similar illness after eating a common food or different food from a common place or (2) an unexplained, unexpected increase of a similar illness, and food is a likely source.

Foodborne illness: A disease acquired through eating or drinking contaminated food or liquids.

Foodborne infection: A disease caused by consuming food or liquids contaminated with bacteria, viruses, or parasites.

Foodborne intoxication: A disease caused by consuming food or liquids contaminated with toxins.

Gastroenteritis: Inflammation of the stomach and intestines.

HACCP[‡]: Hazard Analysis Critical Control Point. A science-based, systematic approach of identifying, evaluating, and controlling food safety hazards.

HACCP plan[‡]: A written documentation of food processing and handling procedures that is based upon the HACCP principles.

HACCP system[‡]: A HACCP plan in operation. The implementation of a HACCP plan.

Hazard: A biological, chemical, or physical agent that may cause foodborne illness.

[‡] FDA. *HACCP: A State-of-the-Art Approach to Food Safety*. http://www.cfsan.fda.gov/~lrd/bghaccp.html.

Highly susceptible population: A group of persons who are more likely than other populations to experience foodborne disease because they are immunocompromised or older adults and in a facility that provides health care or assisted living services, such as a hospital or nursing home; or preschool age children in a facility that provides custodial care, such as a day care center.

Hypothesis: An educated guess based on observations.

Incubation period: The interval from the time an individual is infected to the time when symptoms first appear.

Jaundice: Yellowing of the skin and eyes as a result of accumulation of bile pigment in the blood.

Line listing: A table that summarizes information about persons associated with an outbreak. Information often includes identifying information, demographics, clinical information, and exposure or risk factor information.

Notifiable disease: A disease that is required by law to be reported to the public health authority. See Appendix C for the list of notifiable diseases in Kansas.

Odds ratio (**OR**): The ratio of the odds. This measure of association is used to determine whether a specific exposure is associated with a certain disease.

Onset: The date and time when clinical signs or symptoms first appear.

Outbreak: An unexpected, unexplained increase of disease occurring within a specific population at a given time and place.

Parasite: A single or multi-celled organism that can cause foodborne infections.

Pathogen: A disease-causing organism.

Person-to-person outbreak: See propagated-source outbreak.

Physical hazard*: A foreign material, such as glass, metal, or plastic, that may cause illness or injury

Point-source outbreak: See common-source outbreak.

Potentially Hazardous Food (PHF): Any food or food ingredient (natural or synthetic) that is capable of supporting rapid growth of microorganisms under certain temperatures. Examples include cooked or raw animal products, heat treated vegetables and starches, sprouts, and melons.

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^{*} USDA – FSIS. Guidebook for the Preparation of HACCP Plans. September 1999.

Propagated-source outbreak: A type of outbreak that occurs when infections is spread from one person to another via the fecal-oral route.

Public health surveillance: The routine collection, analysis, summarization, and dissemination of data for the purpose of preventing and controlling the spread of disease.

Pulsed-field gel electrophoresis (PFGE): A laboratory method used to separate bacterial isolates into genetic fragments, thus forming a unique "DNA fingerprint".

p-value: The probability that a difference observed could have occurred by chance alone.

Questionnaire: A predetermined set of questions used to collect data. The main components include identifying information, demographics, clinical information, exposure or risk factor information, and knowledge of illness in others.

Ready-to-eat food: A food item that can be consumed without further preparation. Examples include raw vegetables and fruits, deli meats, bread, and ice.

Recovery period: The period when symptoms decline and illness improves.

Relative risk or relative risk ratio (RR): The ratio of the attack rate for ill persons who were exposed and the attack rate for ill persons who were not exposed.

Reservoir: The source of infection for a susceptible host.

Retail food store: Any establishment or section of an establishment where food and food products are offered to the consumer and intended for off-premises consumption, including delicatessens that offer prepared food in bulk quantities only.

Risk factor: An attribute or exposure that is associated with an increased occurrence of disease or other health-related event or condition.

Spot map: A pictorial of the spatial distribution of illness within a specific setting or area.

Stool: Feces.

Toxin: A poison produced or released by certain bacteria that can cause foodborne intoxications.

Traceback: The method of tracing implicated food items backwards through the production and distribution chain to identify the contaminated item and remove it from the food market.

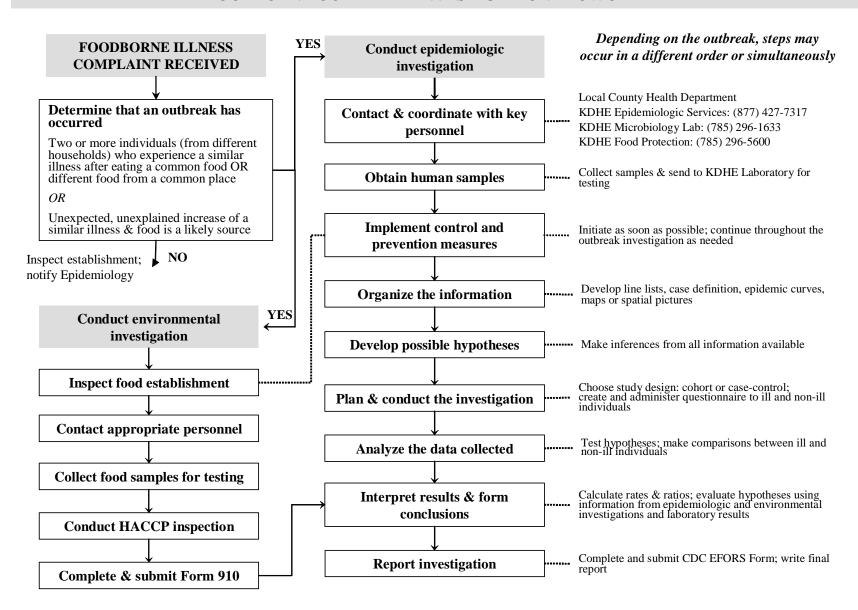
Verification[‡]: Those activities, other than monitoring, that determine the validity of the HACCP plan and that the system is operating according to the plan.

Virus: A minute organism that can cause foodborne infections.

APPENDIX B Flowchart and Checklist

- **B.1** Foodborne Outbreak Investigation Flowchart
- **B.2** Foodborne Disease Outbreak Checklist

FOODBORNE OUTBREAK INVESTIGATION FLOWCHART



Foodborne Disease Outbreak Checklist for Local Health Departments

following checklist provides general steps that infection control nurses or administrators uld take during a foodborne disease outbreak investigation.
Confirm that a foodborne disease outbreak has occurred. Does it meet the definition for a foodborne disease outbreak? Use "Potential Foodborne Outbreak Worksheet" as a helpful tool
Contact and coordinate with KDHE at (877) 427-7317 for all outbreaks.
Contact food inspector, if food service establishment is involved.
Provide stool sample kits to ill persons (5-8 persons) and send samples to KDHE Laboratory. Have stool kits available for distribution Submit human and food samples to KDHE Laboratory using "Universal Form"
 Implement prevention and control measures. Give special restriction or exclusion instructions to ill persons who are food handlers, are associated with day care, or involved with direct patient care. Emphasize good handwashing.
Conduct case finding among other family members, others attending common functions
Continue coordination with KDHE and food inspector assigned to outbreak. • Assist in the administration of questionnaires
If disease is a notifiable disease, report information to KDHE via HAWK, fax, or mail.
Write final report, if resources permit. At the minimum, provide lead investigator with pertinent information to write the final report.

APPENDIX C Forms

- **C.1** Potential Enteric Outbreak Worksheet
- **C.2** Seven-day Enteric Questionnaire
- **C.3** Kansas Notifiable Disease Form
- C.4 CDC "Investigation of a Foodborne Outbreak" Form and Instructions

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POTENTIAL ENTERIC OUTBREAK WORKSHEET

(For use as a tool — submission to KDHE is not necessary)

Date outbreak reported://	Investigator's Initials:						
CALLER INFORMATION							
Name: Agency: City: County: Contact Phone: ()	□ Food Inspector □ Laboratorian □ Nurse □ Physician	Suspected Type of Outbreak Reported: □ Foodborne □ Person-to-person □ Waterborne					
Date First Person Became III:/ Time of First Illness: AM PM	Date of First Known Expo Time of First Exposure:						
# of III Individuals Reported: Total # of Individuals in Group: Symptoms Reported:	□ Day care ˙ □ Nursing Home ˙	□ Private Residence □ Reception Hall □ Restaurant □ School					
 □ Watery Diarrhea □ Bloody Diarrhea □ Fever □ Abdominal cramping □ Headache □ Nausea □ Dizziness □ Other 	Name of Place:						
# of Individuals who: Had doctor visits	Catering Company City & County						
Had ER visits Were hospitalized							
# of Stool Samples Collected To Date (Note: Attempt to collect some samples if none already taken.) COMMENTS							
NEVT STED: Contact KDHE Enido							

NEXT STEP: Contact KDHE Epidemiologic Services of potential outbreak and/or your regional medical investigator at (877) 427-7317 **C.2** Seven-day Enteric Questionnaire

Kansas Department of Health and Environment **7-day Enteric Exposure Questionnaire**

Diagnosis			Date						
Last Name First	st Nam	e	Phone number						
Address		C	City		iteZip code				
Date of birth	Sex_		Occupation						
Enrolled in daycare (children)?	Y	N	Daycare Nan	ne					
First symptom		Date	<i></i>	Time	_ AM	PM			
Indicate all symptoms: Diarrhea (me	ore tha	n 3 loose	e stools in a 24 l	hour period):	Y	N			
Bloody diarr	hea:	Y	N	Number stools/24 h	ours: _				
Stomach ach	ne:	Y	N	Nausea:	Y	N			
Vomiting:		Y	N	Muscle aches/pains	: Y	N			
Fever:		Y	N	Highest temperature	e				
Other sympt	oms?_								
Did you completely recover?	Y	N If so	o: Date:	Time		AM	PM		
Did you see or contact a doctor?	Y	N Nar	ne of doctor						
Was a stool specimen collected?	Y	N We	re you hospitali	zed?	Y	N			
Did anyone else (in your family o	r close	e person	al contacts) ha	ve the same symptor	ns?Y	N			
Relationship:				Date of illness:					
In the 7 days prior to illness, did	you ha	ave cont	act with any of	the following anima	ıls?				
Chicks?	Y	N	Ducklings?	Y N					
Other birds?	Y	N	Specify						
Reptiles (turtles, sna	akes, li	zards, ig	uanas, etc)?	Y N					
Spec	ify								
Other pets or anima	ls?	Y	N Speci	ify					
If any of these animals are									
Store	_			_	<u>.</u>				
Live or work on a farm? Y									
In the 7 days prior to illness, did									

Did you go swimming	in the 7 da	ys pri	or to illness?	Y	N		
If so, did you s	wim in a:	Wa	orinated pool? ding pool? er, Lake, or Pond?	Y Y	N N N		
Name and/or lo	cation of sv	vimmir	ng facility				
Did you travel in the	7 days prio	r to ill	ness?	Y	N		
Specify:	Place				Dates		
1	Place				Dates		
Were you taking anti	biotics at a	nytime	in the 14 days p	rior to	o illness?	Y	N
Name of antibi	otic:			Re	ason for takii	ng antibio	otic:
				_ Date	es taken:		to
Name of antibion	otic:			_ Re	ason for taki	ng antibi	otic:
				Dates taken:		to	
Name/description of ev							Date
What is the source of							
	•	Ü		,	cate all that a		
	Y	N N	Name:				
Bottled Water?		N					
Did you eat at any res	staurants ir	the 7	days before beco	ming	ill?	Y	N
Please list restaurants t	o the best o	f your	recollection:				
1. Name			City		·	Date	
Foods eaten							
2. Name			City			Date	
Foods eaten							
(List additional restaur	ant informa	tion on	reverse)				

(Revised 02/2003)

Indicate which of the below foods you eat routinely.	To the best of your recollection, also provide
the usual brand names and the store name(s) and loca	

Chicken	Y	N	Brand	Store name
				City
Hamburger	Y	N	Brand	
				City
Sausage	Y	N	Brand	
				City
Hot Dogs	Y	N	Brand	
				City
Lunch Meat	Y	N	Brand	
				City
Eggs	Y	N	Brand	
				City
Milk (Raw)	Y	N	Supplier	City
Milk (Past.)	Y	N	Brand	
				City
Fresh Juice	Y	N	Brand	
				City
Fresh Berries	Y	N	Brand	
				City
Fresh Melon	Y	N	Brand	
				City
Other Fresh Fruit	Y	N	Brand	
				City
Lettuce	Y	N	Brand	
				City
Alfalfa Sprouts	Y	N	Brand	
				City
Other fresh vegetab	lesY	N	Brand	Store name
				City
Other foods which	may h	ave cau	used your illness?	
Food Item			Brand	Store name
Food Item			Brand	City Store name
			Drand	City
Any other commen	nts rela	tive to	your illness?	

C.3 Kansas Notifiable Disease Form

KANSAS NOTIFIABLE DISEASE FORM

Today's Date:	.//						
Patient's Name:	Last	First	Middle				
	Last	FIISt	Middle				
Day Phone:		Evening Phone:					
Residential Address	s:						
City:		Zip:	County:				
Ethnicity: Hi	spanic or Latino	Not Hispanic or Latino	Unknown				
	ndian/Alaska Native vaiian or Other Pacific ly)	Asian Islander White	Black or African American Unknown				
Sex: M F	Date of Birth:	//	Age if DOB unknown:				
Disease Name:							
Symptoms: Onset://	State the 3 m	nost prominent symptoms:					
Symptom 1:	Symp	otom 2:	_ Symptom 3:				
Outbreak associated	? Y N	Died? Y	N				
Institutional Reside	ence? None Nu	rsing Home Correctional	l Residential Hospital Psych				
Physician Name:		Physician Phon	e:				
Laboratory Information:							
Specimen Collection	Date:/	/ Date Reported	To You://				
Name of Test Perfor	med:		Results of Test:				
Name of Laboratory: Laboratory Results Attached? Y N							
Treatment Informa	tion:						
Date of Treatment: Treatment Status:	Complete On	Treatment Type and D going Discontinued	Oosage:				
Name of person rep	oorting:		Phone:				
Comments							

C.3

2004 REPORTABLE DISEASES IN KANSAS for health care providers, hospitals, and laboratories (K.S.A. 65-118, 65-128, 65-6001 through 65-6007, K.A.R. 28-1-2, 28-1-4, and 28-1-18)

Bold -- Telephone report within four hours of <u>suspect or confirmed</u> cases to KDHE toll free at 1-877-427-7317.

(i) Isolates must be sent to:

Division of Health and Environmental Laboratories Forbes Field, Building #740, Topeka, KS 66620-0001

Phone: (785) 296-1636

™ DISEASES REQUIRING SPECIAL ATTENTION ™

Anthrax 🕾 Rubella, including congenital rubella syndrome 🕾 Botulism 🕾 Severe Acute Respiratory Syndrome (SARS) (1) 222 Cholera 🕾 Smallpox 🕾 Tuberculosis, active disease (1) 🕾 Measles (rubeola) 🕾 *Meningitis*, bacterial ²²² Viral hemorrhagic fever 🕾 Meningococcemia (1) 222 Escherichia coli O157:H7 Mumps 22 (and other enterohemorrhagic, enteropathogenic and Pertussis (whooping cough) 🕾 enteroinvasive E. coli) ① Plague 🕾 Salmonellosis, including typhoid fever ① Poliomyelitis 🕾 Shigellosis (i) Q Fever 🕾 Streptococcal invasive disease, Group A from Streptococcus or Streptococcus pneumoniae ① Rabies, human and animal 🕾

Acquired Immune Deficiency Syndrome (AIDS)

Amebiasis

Anthrax
Botulism

Brucellosis

Campylobacter infections

Chancroid

Chlamydia trachomatis genital infection

Cholera ≅ Cryptosporidiosis Cyclospora infection Diphtheria Ehrlichiosis

Encephalitis, infectious (includes West Nile virus)

Escherichia coli O157:H7

(and other enterohemorrhagic, enteropathogenic and

enteroinvasive E. coli) ①

Giardiasis Gonorrhea

Haemophilus influenza, invasive disease Hantavirus Pulmonary Syndrome Hemolytic uremic syndrome, postdiarrheal Hepatitis, viral (acute and chronic)

Hepatitis B during pregnancy

Human Immunodeficiency Virus (HIV) (includes Viral

Load Tests) Legionellosis

Leprosy (Hansen disease)

Listeriosis Lyme disease Malaria

Measles (rubeola) 🕾

Meningitis, arboviral (includes West Nile virus)

Mumps 🕾

Pertussis (whooping cough)

Plague Poliomyelitis Posittacosis
Q Fever Positian

Rabies, human and animal ™ Rocky Mountain Spotted Fever

Rubella, including congenital rubella syndrome ≅ Salmonellosis, including typhoid fever ①

Severe Acute Respiratory Syndrome (SARS) ① 🕾

Streptococcal invasive disease, Group A from *Streptococcus* or *Streptococcus pneumoniae* ①

Syphilis, including congenital syphilis

Tetanus

Toxic shock syndrome, streptococcal and staphylococcal

Trichinosis

Tuberculosis, active disease ① [∞] Tuberculosis, latent infection

Tularemia

Yellow fever

In addition, laboratories <u>must</u> report:

- Viral load results of reportable diseases
- ALL blood lead levels, as of 12/2002 (KCLPPP/ABLES)
- CD4+ T-lymphocyte count < 500/ μl or CD4+ T-lymphocytes <29% of total lymphocytes

Outbreaks, unusual occurrence of any disease, exotic or newly recognized diseases, and suspect acts of terrorism should be <u>reported within 4 hours</u> by telephone to the Epidemiology Hotline: <u>1-877-427-7317</u>

C.3

C.4	CDC "Investigation of a Foodborne Outbreak"
	Form and Instructions

FORM APPROVED
OMB NO 0920-0004



 $INVESTIGATION\ OF\ A\ FOODBORNE\ OUTBREAK$ This form is used to report foodborne disease outbreak investigations to CDC. A foodborne outbreak is defined as the occurrence of **two or more cases** of a similar illness resulting from the ingestion of a common food in the United States. This form has **two** parts: Part 1 asks for the minimum data needed and Part 2 asks for additional information. For this investigation to be counted in the CDC annual summary, Part 1 must be completed. We encourage you to complete as much of Part 1 and Part 2 as you can.

OMB NO.0920-0004
CDC USE ONLY
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STATE USE ONLY

Part 1: Paguired Information

		Part 1: Keq	unca	IIIIOIIIIa	11011	-
1. Location of Exposure: State: Multi-state exposure County: Multi-county exposure List other states/counties in Comments, bottom of this page	Date of f	t case became ill: Mo first known exposure:	nth / _ nth / _	Day	Year — — — — Year	3. Numbers of Cases Exposed: Lab-confirmed cases: (A) Probable cases: (B) Estimated total ill: (If greater than sum of A+B)
4. Approximate Percentage of Cases in Each Age Group: <1 year:% 20-49 yrs: 1-4 yrs:% ≥ 50 yrs: 5-19 yrs:%	: %	5. Sex: (Estimated percent of total cases) Male: % Female: %		Interview Case-cor Cohort st	s of cases only ntrol study	(Check all that apply) ☐ Investigation at factory or production plant ☐ Investigation at original source (farm, marine estuary, etc.) ☐ Environment / food sample cultures
7. Implicated Food(s): (based of Reasons listed in Item 15 on page 3		8. Etiology: (Name the type, virulence factors, m Etiology		r fingerprin		f available, include details such as phage metabolic profile. Other Characteristics (if avail.)
□ Could not be determined		□ Suspected □ Pa □ Fo □ Unknown etiology □ En			d from (check all that apply) Patient specimen(s) Food specimen(s) Environment specimen(s) Food Worker specimen(s) 2000/Vol 49/SS-1/Appendix B	
9. Contributing Factors: (See Contributing factors unknown Contamination Factor: C1 C2 C3 C3 C10 C11 C12 C3 C10 C11 C12 C6 C10 C11 C12 C7 C10 C11 C12 C12 C12 C11 C12 C12 C12 C12 C13 C12 C13 C13 C13 C13 C13 C13 C13 C13 C13 C14 C15	n C4	C5	□ C8	□ P9	Contact Per NAME: TITLE: PHONE NO: FAX NO: E-MAIL: Date of com	
Was food-worker implicated as the source of contamination? ☐ Yes ☐ No If yes, please check <i>only one</i> of following: ☐ laboratory <i>and</i> epidemiologic evidence ☐ epidemiologic evidence (w/o lab confirmation) ☐ lab evidence (w/o epidemiologic confirmation) ☐ prior experience makes this the likely source (please explain in Comments) ☐ Initial Report ☐ Updated Report ☐ Final Report ☐ Additional data suggests this is not a foodborne outbreak						

Comments:

This questionnaire is authorized by law (Public Health Service Act, 42 USC §241). Although response to the questions asked is voluntary, cooperation of the patient is necessary for the study and control of disease. Public reporting burden for this collection of information is estimated to average 15 minutes per response. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden to PHS Reports Clearance Officer; Rm 721-H, Humphrey Bg; 200 Independence Ave. SW; Washington, DC 20201; ATTN: PRA, and to the Office of Information and Regulatory Affairs, Office of Management and Budget, Washington, DC 20503.

The following codes are to be used to fill out Part 1 (question 9) and Part 2 (question 15).

Contamination Factors:1

- C1 Toxic substance part of tissue (e.g., ciguatera)
- C2 Poisonous substance intentionally added (e.g., cyanide or phenolphthalein added to cause illness)
- C3 Poisonous or physical substance accidentally/incidentally added (e.g., sanitizer or cleaning compound)
- C4 Addition of excessive quantities of ingredients that are toxic under these situations (e.g., niacin poisoning in bread)
- C5 Toxic container or pipelines (e.g., galvanized containers with acid food, copper pipe with carbonated beverages)
- C6 Raw product/ingredient contaminated by pathogens from animal or environment (e.g., Salmonella enteriditis in egg, Norwalk in shellfish, E. coli in sprouts)
- C7 Ingestion of contaminated raw products (e.g., raw shellfish, produce, eggs)
- C8 Obtaining foods from polluted sources (e.g., shellfish)
- C9 Cross-contamination from raw ingredient of animal origin (e.g., raw poultry on the cutting board)
- C10 Bare-handed contact by handler/worker/preparer (e.g., with ready-to-eat food)
- C11 Glove-handed contact by handler/worker/preparer (e.g., with ready-to-eat food)
- C12 Handling by an infected person or carrier of pathogen (e.g., Staphylococcus, Salmonella, Norwalk agent)
- C13 Inadequate cleaning of processing/preparation equipment/utensils leads to contamination of vehicle (e.g., cutting boards)
- C14 Storage in contaminated environment leads to contamination of vehicle (e.g., store room, refrigerator)
- C15 Other source of contamination (please describe in Comments)

Proliferation/Amplification Factors:1

- P1 Allowing foods to remain at room or warm outdoor temperature for several hours (e.g., during preparation or holding for service)
- P2 Slow cooling (e.g., deep containers or large roasts)
- P3 Inadequate cold-holding temperatures (e.g., refrigerator inadequate/not working, iced holding inadequate)
- P4 Preparing foods a half day or more before serving (e.g., banquet preparation a day in advance)
- P5 Prolonged cold storage for several weeks (e.g., permits slow growth of psychrophilic pathogens)
- P6 Insufficient time and/or temperature during hot holding (e.g., malfunctioning equipment, too large a mass of food)
- P7 Insufficient acidification (e.g., home canned foods)
- P8 Insufficiently low water activity (e.g., smoked/salted fish)
- P9 Inadequate thawing of frozen products (e.g., room thawing)
- P10 Anaerobic packaging/Modified atmosphere (e.g., vacuum packed fish, salad in gas flushed bag)
- P11 Inadequate fermentation (e.g., processed meat, cheese)
- P12 Other situations that promote or allow microbial growth or toxic production (please describe in Comments)

Survival Factors:1

- S1 Insufficient time and/or temperature during initial cooking/heat processing (e.g., roasted meats/poultry, canned foods, pasteurization)
 - S2 Insufficient time and/or temperature during reheating (e.g., sauces, roasts)
 - S3 Inadequate acidification (e.g., mayonnaise, tomatoes canned)
 - S4 Insufficient thawing, followed by insufficient cooking (e.g., frozen turkey)
 - S5 Other process failures that permit the agent to survive (please describe in Comments)

Method of Preparation:²

- M1 Foods eaten raw or lightly cooked (e.g., hard shell clams, sunny side up eggs)
- M2 Solid masses of potentially hazardous foods (e.g., casseroles, lasagna, stuffing)
- M3 Multiple foods (e.g., smorgasbord, buffet)
- M4 Cook/serve foods (e.g., steak, fish fillet)
- M5 Natural toxicant (e.g., poisonous mushrooms, paralytic shellfish poisoning)
- M6 Roasted meat/poultry (e.g., roast beef, roast turkey)
- M7 Salads prepared with one or more cooked ingredients (e.g., macaroni, potato, tuna)
- M8 Liquid or semi-solid mixtures of potentially hazardous foods (e.g., gravy, chili, sauce)
- M9 Chemical contamination (e.g., heavy metal, pesticide)
- M10 Baked goods (e.g., pies, eclairs)
- M11 Commercially processed foods (e.g., canned fruits and vegetables, ice cream)
- M12 Sandwiches (e.g., hot dog, hamburger, Monte Cristo)
- M13 Beverages (e.g., carbonated and non-carbonated, milk)
- M14 Salads with raw ingredients (e.g., green salad, fruit salad)
- M15 Other, does not fit into above categories (please describe in Comments)
- M16 Unknown, vehicle was not identified

2 of 6

¹ Frank L. Bryan, John J. Guzewich, and Ewen C. D. Todd. Surveillance of Foodborne Disease III. Summary and Presentation of Data on Vehicles and Contributory Factors; Their Value and Limitations. Journal of Food Protection, 60; 6:701-714, 1997.

² Weingold, S. E., Guzewich JJ, and Fudala JK. Use of foodborne disease data for HACCP risk assessment. Journal of Food Protector, 57; 9:820-830, 1994.

	Part 2: Ad	ditional Information	(Pl	ease con	nplete as mu	ich as	s possible)		
11. Numbers of: OUTCOME / SYMPTOM	Cases with Outcome / Symptom	Total cases for whom you have information available		12. Incubation Periodic (circle appropriate appropriat		ınits)	13. Duration of Acute Illness Among Those Who Recovered: (circle appropriate units)		
Healthcare Provider Visit	- J		Sh	ortest:	` ′	• .			_ (Hours, days)
				ngest:	•	• .			_ (Hours, days)
Hospitalization			IVIE	edian:	(Hours, d	ays)			_ (Hours, days)
Death			□ Unknown		□ Unknown				
Vomiting									
Diarrhea						ppropr	riate, to describe o	ther	common
Bloody stools			cna	aracteristics	s of cases:				
Feverish				anaphyla: arthralgia		cendir hing	0. ,	nyal bare:	gia sthesia
Abdominal cramps				bradycard	dia hea	adache		septi	icemia
*				bullous sl lesions		•			throat ycardia
*				bradycard cough		otensi ing			nobocytopenia perature reversal
*				coma	jau	ndice	į	urtica	aria
*				diplopia	ieu	argy	\	MILE	ezing
14. If Cohort Investigat	ion Conducte	ed:							
Event-specific Attack F	Rate =	1					x 10	0 =	=%
		# ill tot	al#c	of persons fo	or whom you hav	e illnes	ss info.		
15. Implicated Food(s):	(Please prov	vide known information.)			Po	eason(s) Suspected		Method of Preparation
Name of Food	Main Ingredier	its				e below) (see list on page 2)			
e.g., lasagna	pasta, sauc	e, eggs, beef		eggs 4		4	4		M1
☐ Food vehicle could not be	e determined								
	ence from epidem dence (e.g., identi	niological investigation fication of agent in food)					found on farm that si experience makes th		
16. Where was Food P	repared? (Ch	eck all that apply)			17. Where v	vas F	ood Eaten? (Ch	neck	call that apply)
□ Restaurant or deli □ Prison, jail □ Day care center □ Private home □ School □ Picnic □ Church, temple, etc. □ Camp □ Contaminated food imported into U.S. □ Caterer □ Commercial product, served without for preparation □ Other (please describe) □ Workplace cafeteria □ Nursing home □ Prison, jail □ Private home □ Privat			S.		Restaurant or deli Day care center School Church, temple, etc. Camp Grocery Store Hospital Workplace cafeteria Nursing home Prison, jail Private home Fair, festival, or m location Other (please des		, jail e home estival, or mobile on		
18. Other Available Info ☐ Unpublished agency rep (please attach) ☐ Epi-Aid ☐ Publication (please refer	19. Remarks: Bri (e.g., restaurant	-		= =				covered above nomic impact, etc.)	

 $State\ Health\ Departments:\ Please\ FAX\ this\ document\ to\ Foodborne\ and\ Diarrheal\ Diseases,\ DBMD,\ CDC,\ at\ (404)\ 639-22054$

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Guidelines for completing the form "Investigation of a Foodborne Outbreak" Foodborne and Diarrheal Diseases Branch, DBMD, CDC November 20, 2000

1. Location of Exposure

Provide two-letter postal code of the state, and the full name of the county in which <u>exposure</u> took place. If exposure took place in multiple states or counties (such as with a commercial product), check the box provided and list other states or counties in the Comments section at the bottom of page 1.

2. Dates

Indicate date that first known case patient became ill, and date that the first and last known exposure took place. If available, please send a copy of the epidemic curve along with this report form.

3. Numbers Exposed in Your Jurisdiction

Provide number of laboratory-confirmed cases and number of presumptive cases. If applicable, also provide an estimate of the total number of ill persons if you suspect that this number exceeds the sum of the laboratory-confirmed and presumptive cases.

4. Approximate Percentage of Total Cases in Each Age Group

This item seeks to identify unique patterns of age distribution for the outbreak, as well as to identify age groups most affected. Indicate the approximate percentage of all cases (lab-confirmed and presumptive) in the various age groups listed. Total should equal 100%.

5. Sex

Estimate the percentage of males and females, using all cases (laboratory-confirmed and presumptive combined). Total should equal 100%.

6. Investigation Methods

Check off all boxes that describe the methods used to investigate this outbreak.

7. *Implicated Food(s)*

List the food item(s) implicated as a result of the investigation. Response to this question should match response to Item 15 on page 3, and should be based on one of the <u>Reasons Suspected</u> given in Item 15.

8. Etiology

- A. Identify the bacterium, virus, parasite, or toxin responsible for the outbreak. Please give as much detail as you have about the organism or toxin.
- B. Check the box to indicate whether the etiology is confirmed or suspected. "Confirmed" means that the criteria for confirmation of that etiology have been met. (Please visit our website at http://www.cdc.gov/ncidod/dbmd/outbreak/guide_fd.htm or see MMWR 2000 / Vol. 49 / ss-1 / Appendix B for confirmation criteria...).
- C. If more than one etiology was identified, please describe in the Comments section at the bottom of page 1.
- D. Check off all boxes that correspond to the specimen(s) from which the etiologic agent was

isolated or identified.

9. Contributing Factors

- A. Factors that contribute to the occurrence of outbreaks are classified according to contamination, survival, and proliferation. A factor should be checked only if the investigator has strong evidence that it actually occurred in this outbreak; just because a factor has been cited in similar outbreaks in the past does not mean it was involved in this outbreak. Contamination factors relate to how the agent got onto or into the food vehicle. Proliferation factors relate to how microbial agents were able to increase in numbers and/or produce toxic products prior to the vehicle being ingested. Survival factors refer to processes or steps that should have eliminated or reduced the agent but did not for the reason listed. Explanations and examples of the codes are provided on page 2 of the form. If the choice of "other" is made for any of the factors, please describe in the Comments section at the bottom of page 1.
- B. If one or more food workers are implicated as the source of contamination, please indicate what evidence was used to support this conclusion. The choice of "prior experience makes this the likely source" is provided for situations when conclusive laboratory and epidemiologic evidence is absent, but other factors may prompt the investigator to suspect the food worker(s). If a food worker is implicated in the absence of laboratory and/or epidemiologic evidence, please explain in the Comments section at the bottom of page 1.

10. Agency reporting this outbreak

Indicate the agency reporting the outbreak and the relevant information for the person to contact with questions regarding the outbreak investigation. Provide today's date, and indicate whether this is the initial report of the outbreak investigation, or an update to a prior report.

11. Numbers

For each outcome listed, provide the number of patients with the outcome, and the total number of patients for whom you have such outcome information available. If applicable, list other outcomes (and the relevant numbers) in the blank spaces provided. A list of possible outcomes is provided to the right of the table.

12. Incubation Period

Indicate the shortest, longest, and median incubation period, and indicate whether each period is measured in hours or days.

13. Duration of Acute Illness Among Those Who Recovered Indicate the shortest, longest, and median duration of acute illness among those who recovered. Indicate whether each period is measured in hours or days.

14. If Cohort Investigation Conducted

For cohort investigations only, report the attack rate. The formula is provided to aid in keeping our definition of attack rates consistent across investigations.

15. *Implicated Food(s)*

Foods implicated in outbreaks may contain multiple ingredients, while often only one of these ingredients is the actual source of the etiologic agent. When possible, identification of an implicated ingredient(s) provide a basis for identifying ingredients that may be involved in other outbreaks. Please list the name of the food, the main ingredients, the contaminated ingredient, and the reason(s) for suspecting that the particular ingredient(s) was contaminated. Indicate method of preparation using list of codes found on page 2 of the form.

16. Where was Food Prepared?

Indicate where food was prepared. Check all applicable boxes.

17. Where was Food Eaten?

Indicate where suspected/implicated food was eaten. Check all applicable boxes.

18. Other Available Information

Indicate what other sources of information are available for this outbreak. References should be cited for published papers.

19. Remarks

Describe other important aspects of the outbreak that may not have been reported elsewhere in the form.

State health departments should fax form to the Foodborne and Diarrheal Diseases Branch, CDC, at 404-639-2205, or mail to: FBO Reporting, Foodborne and Diarrheal Diseases Branch, CDC Mail Stop A-38, 1600 Clifton Road, Atlanta, GA, 30333.

APPENDIX D Sample Collection

- **D.1** General Guidelines for Stool Sample Collection
- **D.2** General Guidelines for Food Sample Collection
- **D.3** Requisition Form for Ordering Stool Kits
- D.4 Sample Copy of KDHE Universal Laboratory Specimen Submission Form

D.1	General Guidelines for Stool Sample Collection

GENERAL GUIDELINES FOR STOOL SAMPLE COLLECTION

The KDHE laboratory provides two types of stool kits to LHDs:

- (1) Enteric kits, which test for *Campylobacter*, *Salmonella*, *Shigella*, and virulent *E. coli* for culture and identification. These kits may also be used to test for Norovirus. However, testing for Norovirus is conducted only in outbreak situations and must be cleared by Epidemiologic Services at (877) 427-7317.
- (2) Ova and parasite (O&P) kits, which identify intestinal parasites, including *Cryptosporidium* parvum and *Giardia lamblia*.

Three sets of each kit (enteric and O&P) are recommended to be kept on hand.

Requesting stool kits from the KDHE laboratory:

- 1. Complete the following requisition for laboratory specimen kits.
- 2. Under the subheading Bacterial Culture, indicate the number of kits requested next to the label "Enteric".
- 3. Under the subheading Parasite (O&P), indicate the number of kits requested next to the label "Feces".
- 4. Fax completed form to the KDHE laboratory at (785) 296-1641.

Distributing and Collecting Stool Samples:

- 1. Provide 1 stool kit to persons experiencing diarrhea, defined as 3 or more loose stools within a 24-hour period. In outbreak situations, 5 to 8 stool samples are ideal.
- 2. Ask person(s) to return stool sample(s) to health department.
- 3. Fill out all items on Page 1 of the KDHE Laboratory Universal Form.
- 4. If bacterial cultures are requested, mark the appropriate box on Page 2, under the subheading Bacteriology Culture.
- 5. If parasitic identification is requested, mark the appropriate box on Page 2, under the subheading Parasitology.
- 6. Apply one barcode label from the KDHE Laboratory Universal Form to each stool collection vial.
- 7. Place vials and universal form in an appropriate shipping container.
- 8. Mail stool sample(s) to the following address:

Kansas Department of Health and Environment

Division of Health & Environmental Laboratories

Forbes Field, Building 740

Topeka, KS 66620

For questions regarding patients and symptoms, contact Epidemiologic Services at (877) 427-7317. For questions regarding laboratory specimen submission, contact the KDHE Laboratory at (785) 296-1620.

D.2	General Guidelines for Food Sample Collection

GENERAL GUIDELINES FOR FOOD SAMPLE COLLECTION

Microbiological analysis of food is provided in support of an investigation of a foodborne disease outbreak. The purpose of testing is to isolate and identify pathogenic microorganisms in food samples, which have been implicated in the outbreak.

Submission of Food Samples for Analysis

Food sample collection is most often conducted by a food inspector. However, submission to the KDHE laboratory for testing should be conducted by the local health department (LHD).

All requests for laboratory examination of food or food-related samples must be made through KDHE's Epidemiologic Services at (877) 427-7317. Laboratory examination of food will be performed only when pre-approved. Results of food analysis will be reported to Epidemiologic Services and the LHD responsible for submission.

Method for Collecting Aseptic Samples

The value of laboratory results in microbiology depends on the quality of the samples submitted. Food samples must be obtained using aseptic technique and appropriate containers. Samples must be refrigerated during storage and transport and must arrive at the food microbiology laboratory within three days of collection. Samples collected frozen should be stored and transported frozen (on dry ice).

- 1. Use sterile containers and ensure that leakage is prevented.
- 2. Wash hands prior to putting gloves on.
- 3. Do not touch inside of collection container.
- 4. Use sterile utensils, such as a tongue depressor to collect food.
- 5. Collect an adequate amount of the sample—a minimum of 4-6 ounces, if possible.
- 6. Fill the container no more than 3/4 full.
- 7. Keep food cold by placing it in the styrofoam cooler with an ice pack. Use additional ice when necessary.
- 8. Clearly document how the product was handled and who handled it after the sample is taken.

Labeling Food Samples

- 1. Name and type of product
- 2. Brand of product
- 3. Product manufacturer
- 4. Inspector name
- 5. Date, time, and place of collection
- 6. Establishment name

Transporting Food Samples

- 1. Submit sample to the LHD infection control nurse in the county where the outbreak occurred.
- 2. LHD nurse should complete a KDHE Universal Laboratory Specimen Submission Form. On Page 1, under "Sample Information" and subheading "Clinical Source", mark "Other" and write 'Food Sample'. On Page 2, under the subheading "Submitter Comments", indicate that the food is related to an outbreak investigation and that testing was approved by KDHE Epidemiology at (877) 427-7318.
- 3. Sample should be sent via the KDHE courier or FedEx using ice packs and an insulated container.

NOTE: Food should be kept cold during transportation and placed in a refrigerator when not in transit.

Recommended Sampling Equipment

- Sterile sample containers
 - o Plastic bags (Whirl-Pak)
- Sterile and wrapped sample collection implements
 - o Tongue depressors
 - o Swabs
 - o Individually wrapped plastic spoons
- Supporting equipment
 - o Individually wrapped disposable gloves
- Sterilizing and sanitizing agents
 - Alcohol swabs
- Refrigerants
 - o Ice pack
 - o Thermometer
 - o Insulated container (Styrofoam cooler)

D.3	Requisition Form for Ordering Stool Kits



DIVISION OF HEALTH AND ENVIRONMENTAL LABORATORIES DEPARTMENT OF HEALTH AND ENVIRONMENT

Forbes Field, Building 740 Topeka, Kansas 66620-0001

REQUISITION FOR LABORATORY SPECIMEN KITS

Please use the appropriate kit listed below to submit specimens to the Health and Environmental Laboratories. Each kit consists of a specimen container, an addressed mailing container, and a test request form. If you have any questions about submitting specimens, please refer to the Manual of Laboratory Tests or call (785) 296-1623.

RUSH ORDERS: FAX to (785) 296-1641

Please enter the quantity needed on the line next to the item.

Serology	Parasite (O & P)
Single Mailer	Feces
Multi-mailer without box	Pinworm*
Multi-mailer with box	<u>Gonorrhea</u>
<u>Viral Culture</u>	Culture Plates
Virus*	Mailer, Two Specimen
Neonatal Screening	CO ₂ Tablets
Initial (Green) Collection Unit	Whirl-Pak Bag
Repeat (Red) Collection Unit	Bacterial Culture
Inorganic Chemistry	Enteric
Filter Paper Forms for Blood Lead	TB, Sputum
Blood Lead Confirmation Kits	TB, Gastric
	Miscellaneous - Infectious Substance Shipper
Forms Only	Miscellaneous Kit for Non-Infectious Specimens
Universal	<u>Other</u>
Blood Lead Confirmation	(Specify):
Serology Assays: Hepatitis (HBsAg*	*, HAV-IgM***, HCV****), Rubella*, HIV **
Provided to City/County Health Departments only. *** Contact Epidemiologic Services 296-1127 first .	** AIDS Counseling & Testing and Prenatal Testing sites only *** Drug abuse associated HIV CT Clients only.
Send to:	
Facility ID No. :	_
Facility Name:	LAB USE ONLY
Attn:	Order Number
Address:	Date Received
City:, KS	D.
Phone:	-

D.4	Sample Copy of KDHE Universal Laboratory
	Specimen Submission Form

KDHE Universal Laboratory Spe	ecimen Submission Form (Health)	Page 1
2 Use only pens with dark ink. 3 Fill in squares LIKE THIS:	letters only and completely inside boxes: C 1 2 3 plete all items on form.	
NOTE: THE ENTIRE LABEL MUST BE APPLIED COLLECTION TUBE. ENTIRE BARCODE MUST IN NABLE LENGTHWISE! (SEE DIAGRAM BELOW	BE VISIBLE AND SC AIN	
123456	1 1 2 3 4 5	6_71_2_3_4_6_6_7
PROVIDER INFORMATION:		1
FACILITY ID PHYSICIAN'S	T NAI.	
PATIENT INFORMATION:		
PATIENT'S LAST NAME		
PATIENT'S FIRST NAME	KDHE App (5) Affix 4" x 1" Label Call (785) 296-16	Ved Label Only Uprion and Level!! Stor Information
PATIENT'S CODE	SEX BIRTOPATE	CO OF RES
	_ M _ F	/
MEDICAIC NUMBER	RACE	ETHNICITY
	Wks Asia AI, AN	Hispanic or Latino Non-Hispanic or Non-Latino
SAMPLE INFORMATION:	6, 90,	
DATE OF COLLECTION	DATE OF ONSE	PATIENT SYMPTOMS Yes No
CLINIC SOURCE	SPECIMAN TYPE	
Adolescent Prison C & T University Drug STD FP TB M & I Other* Prenatal *Specify	Serum Ui CSF Sputum Va Endocervical Stool W Genital Throat O	rine Pericardial Fluid rethral Peritoneal Fluid aginal Pleural Fluid found Synovial Fluid ther* Thoracentesis Fluid
ACUTE SERUM	CONV	ALESCENT SERUM
		/ / /
DO NOT WRITE IN THE SPACE	CE BELOW - DO NOT PHOTO	COPY THIS FORM
Kansas Department of Health and Environment Division of Health & Environmental Laboratories Forbes Field, Building 740, Topeka, KS 66620 CLIA #17DO648254 Phone (785) 296-1620 Fax (785) 296-1641 V03.1	1 2 3 4 5	D.4

TEST INFORMATION/REC	QUEST	
HIV Serology	· · · · · · · · · · · · · · · · · · ·	-
Risk Code Ref Code Prior Confirm	ation Specimen	est Purpose
Yes	Initial Then Referral	Diagnosis Other
No No	Repeat	Prenatal
Hepatitis If a HBsAG is requested with and blood must be submitted (HAV, F	other serology text 5 m of serum of tubes of Other S	Serological Assays
HAV-IgM Exposure Risk	HCV-lqG VDU Hist V/Sexual Partner	Vaccine Preventable
HBsAG Household Contact & Prenatal	IgG CSF	Other Specify
Sexual Contact	Q tel Assays	Сроспу
Syphilis Serology Test Purpose Clinical Information	Rrjor Reagin Reactive Test	Rubella
Diagnosis Asymptomatic	RPR, RST or VDRL	Immune Status/Prenatal
Prenatal Late Syphilis Syntton	Test Date 1)	Diagnosis
Other Treatment otro	2)	Date of Exposure
Nucleic Acid Amplified	Namydia and Gonorrhea	Pertussis
Exam Purpose Clinica Obs	vations Risk History	
Comp FP Exam	Friable New Partner None	PCR
PN Exam Repeat S-Like	Multiple Partners None Contact of STD Case	Other
	_ none	
Viral Cultures Specimen aterial	Viral Syndrome Observed	
Body Flui	Gastroenteritis Ocular	Vesicles
Compsy	Genital Lesion Respirat	
Autopsy	☐ Vaccine Preventable Disease ☐ N ☐ Other - Specify	al acify
		
Blood Lead Patie	nt Address Required for Blood Lead Specimens	
Capillary Patient Address		
Venous		
Repeat Specimen City, State, Zip		
Bacteriology Culture	Parasitology	
	Farasitology	
Enteric Screen R/O Other Enteric Organisms	Intestinal Parasite (Not Cryptosparidium)	Non-Fecal Specimen
Specify	D/O Crysta.	Spacify.
Bacterial Identification		Specify
	should include one of regions):	
Suspected Gonorrhea Culture		
Suspected	Watery Dian ea	Arthropod/Insect ID
Suspected Gonorrhea Culture	Watery Dial ea Institution Re. dent	Arthropod/Insect ID Pinworm Exam (Co. Health Dept. Only)
Suspected Gonorrhea Culture (non-genital/legal) Tuberculosis Culture w/Smear	Watery Dial ea Institution Re. dent	
Suspected Gonorrhea Culture (non-genital/legal) Tuberculosis	Watery Dial ea Institution Re. dent Immune Suppressed	
Suspected Gonorrhea Culture (non-genital/legal) Tuberculosis Culture w/Smear Mycobacterium Isolate for ID	Watery Dial ea Institution Re. dent Immune Suppressed	
Suspected Gonorrhea Culture (non-genital/legal) Tuberculosis Culture w/Smear Mycobacterium Isolate for ID	Watery Dial sa Institution Re. dent Immune Supproved	
Suspected Gonorrhea Culture (non-genital/legal) Tuberculosis Culture w/Smear Mycobacterium Isolate for ID CDC Provided Tests Subm	Watery Dial sa Institution Re. dent Immune Supproved	
Suspected Gonorrhea Culture (non-genital/legal) Tuberculosis Culture w/Smear Mycobacterium Isolate for ID CDC Provided Tests Specify Subm	Watery Dial sa Institution Re. dent Immune Supproved	Pinworm Exam (Co. Health Dept. Only)

Kansas Department of Health and Environment

Kansas Department of Health and Environment Division of Health & Environmental Laboratories Forbes Field, Building 740, Topeka, KS 66620 CLIA #17DO648254 Phone (785) 296-1620 Fax (785) 296-1641 V03.1



APPENDIX E Data Analyses

E.1 Conducting Data Analyses

ANALYZING THE DATA COLLECTED

The following should be calculated to understand the data collected: frequencies and percentages, the incubation and recovery periods, the measures of association between exposure and disease, and appropriate tests of statistical significance.

EpiInfoTM Version 3.2.2 (released April 14, 2004) is a user-friendly, Windows-based software that is available for free by the CDC that may be used to conduct these specific analyses. Visit www.cdc.gov/epiinfo/ to download.

STEP 1. Calculate frequencies

COUNTS and **PERCENTAGES** should be calculated to define the outbreak. The following is a list of common calculations.

- o For all individuals (cohort or case-control study)
 - Number and percentage of persons by sex
 - Median age and age range
 - Number and percentage of ill ("cases") and not ill ("controls")
 - Other characteristics may be recommended depending upon the outbreak, including race/ethnicity, occupation, county, state, zip code, school name, school grade, nursing home name, room number
- o Total number of persons exposed (cohort study)
- o For ill persons or cases (cohort or case-control study)
 - Number and percentage of each symptom experienced
 - Diarrhea (3 or more loose stools in a 24-hour period), bloody diarrhea, vomiting, nausea, abdominal cramping, fever, malaise, headache are common symptoms
 - Number of samples collected and submitted for testing
 - Stool, blood, urine
 - Number and percentage of laboratory-confirmed results
 - Number and percentage of persons hospitalized
 - Number and percentage of medical visits

STEP 2. Calculate the incubation period and recovery period

The **INCUBATION PERIOD** is the interval from the time an individual is infected (exposed) to the time when symptoms first appear. The incubation period may differ from person to person and from organism to organism.

Incubation period = onset time – time of exposure

EXAMPLE: Time of exposure = 8:00 P.M. on Saturday evening Onset of symptoms = 2:00 A.M. on Monday morning

1. Determine how many hours there were per day.

Hint: Time listed in military time and days listed in 24-hour increments will make calculations much easier for most analytic software.

<u>Saturday:</u> 8:00 P.M. is equivalent to 20:00 in military time. So, 24:00 - 20:00 = 4 hours

Sunday: All hours were of interest = 24 hours

Monday: 2:00 A.M. is equivalent to 02:00 in military time = 2 hours

2. Add all the hours together for the days of interest.

$$4 + 24 + 2 = 30$$
 hours

30 hours / 24 hours = 1.25 days

Incubation period for ill person = 30 hours or 1.3 days

3. Calculate the incubation period for each ill person. Determine the **median** incubation period (the mid-point or middle value) and the **range** (the minimum and maximum numbers).

Person	Incubation Period			
# 1	14 hours	←	Minimum	In out of on movied
# 2	18 hours			Incubation period
# 3	23 hours			median = 30 hours
# 4	30 hours	\leftarrow	Median	T
# 5	36 hours			Incubation period range
# 6	45 hours			= 14 hours - 47 hours
# 7	47 hours	\leftarrow	Maximum	

The **RECOVERY PERIOD** is the period when symptoms decline and illness improves.

Recovery period = recovery time – onset time

EXAMPLE: Onset of symptoms = 2:00 A.M. on Monday morning Recovery from illness = 11:00 P.M. on Tuesday evening

1. Determine how many hours there were per day.

Hint: Time listed in military time and days listed in 24-hour increments will make calculations much easier for most analytic software.

Monday: 2:00 A.M. is equivalent to 02:00 in military time. So, 24:00 - 2:00 = 22 hours Tuesday: 11:00 P.M. is equivalent to 23:00 in military time. So the hours of interest on Tuesday = 23 hours

2. Add all the hours together for the days of interest.

$$22 + 23 = 45$$
 hours

$$45 \text{ hours} / 24 \text{ hours} = 1.88 \text{ days}$$

Recovery period for ill person = 45 hours or 1.9 days

3. Calculate the recovery period for each ill person. Determine the **median** recovery period (the mid-point or middle value) and the **range** (the minimum and maximum numbers).

<u>Person</u>	Recovery Period			
# 1	28 hours	\leftarrow	Minimum	D
# 2	29 hours			Recovery period median
# 3	33 hours			= 37 hours
# 4	37 hours	\leftarrow	Median	D
# 5	40 hours			Recovery period range
# 6	45 hours			= 28 hours - 51 hours
# 7	51 hours	\leftarrow	Maximum	

STEP 3. Utilize the 2 x 2 table to calculate measures of association

A 2 x 2 contingency table can be used to compare the association between illness and exposure.

	Ill	Not Ill	
Exposed	a	b	$(\mathbf{a} + \mathbf{b})$
Not Exposed	c	d	$(\mathbf{c} + \mathbf{d})$
	$(\mathbf{a} + \mathbf{c})$	$(\mathbf{b} + \mathbf{d})$	a+b+c+d

Interpretation of the elements in the 2 x 2 table

 \mathbf{a} = the number of ill persons who were exposed to a specific risk factor

 \mathbf{b} = the number of persons who did not become ill, but were exposed to a specific risk factor

 \mathbf{c} = the number of ill persons who were not exposed to a specific risk factor

 \mathbf{d} = the number of persons who did not become ill and were not exposed to a specific risk factor

 $(\mathbf{a} + \mathbf{b})$ = the total number of persons exposed

(c + d) = the total number of persons not exposed

 $(\mathbf{a} + \mathbf{c})$ = the total number of ill persons

 $(\mathbf{b} + \mathbf{d})$ = the total number of not-ill persons

 $\mathbf{a} + \mathbf{b} + \mathbf{c} + \mathbf{d} =$ the total number of persons

CALCULATIONS FOR COHORT STUDIES

Using a 2 x 2 table, attack rates, food-specific attack rates, and relative risk ratios may be calculated to describe the association between illness and exposure for cohort studies.

ATTACK RATES (includes food-specific attack rates)

An **attack rate** represents the occurrence of disease observed among a defined population over a limited period of time. Specifically, it is used to calculate (1) the percentage of illness among all individuals who were exposed to a specific risk factor and (2) the percentage of illness among all individuals who were not exposed to the specific risk factor.

Attack rate for ill persons who were exposed =
$$\frac{a}{a+b}$$
 X 100

Attack rate for ill persons who were not exposed =
$$\frac{c}{c+d}$$
 X 100

RELATIVE RISK

A **relative risk** (**RR**) is the measure of association between exposure and illness used for cohort studies. It is the ratio of the attack rate for ill persons who were exposed and the attack rate for ill persons who were not exposed.

Relative risk ratio =
$$\frac{\text{Attack rate for ill persons who were exposed}}{\text{Attack rate for ill persons who were not exposed}} = \frac{a/a+b}{c/c+d}$$

Interpretation of Relative Risk (RR)[†]:

 $\mathbf{RR} = \mathbf{1}$: The risk of illness among exposed persons is the same as the risk of illness among those not exposed.

RR>1: The risk of illness among exposed persons is higher than the risk of illness among those not exposed.

RR<1: The risk of illness among exposed persons is lower than the risk of illness among those not exposed.

[†] "E. coli O157:H7 Infection in Michigan" Computer-based Case Study. <u>www.phppo.cdc.gov/phtn/casestudies/computerbased</u>.

EXAMPLE: One hundred fifty individuals attended a wedding reception. Several persons became ill with diarrhea and vomiting between 12 and 48 hours after eating food served at the reception. Calculate the attack rates for (1) ill persons who ate the food served at the reception and (2) ill persons who did not eat the food served at the reception. Also calculate the relative risk (RR) ratio and interpret the results.

	III	Not Ill	
Ate food at reception	72	63	135
Did not eat food at reception	2	13	15
	74	76	150

Attack rate for ill persons who ate food =
$$\frac{72}{135}$$
 X 100 = 53.3%

Attack rate for ill persons who did not eat food =
$$\frac{2}{15}$$
 X 100 = 13.3%

RR = Attack rate for ill persons who ate at the reception
Attack rate for ill persons who did not eat at the reception
$$= \frac{.533}{.133} = 4.0$$

Interpretation: About 53% of the persons who became ill had eaten the food served at the reception compared to 13% who became ill and had not eaten the food. In other words, the risk of illness among persons who ate food at the reception appears to be 4 times higher than the risk of illness among persons who did not eat food at the reception.

EXAMPLE: One hundred thirty-five individuals attended the wedding reception and ate the food served. Specific foods served included salad, dinner rolls, chicken, and cake. Calculate the food-specific attack rates for (1) ill persons who ate each of these items and (2) ill persons who did not eat each of these items. Also calculate the respective relative risk (RR) ratios and interpret the results.

EATING SALAD vs. BECOMING ILL

_	Ill	Not Ill	
Ate salad	63	52	115
Did not eat salad	10	10	20
•	72	63	1 35

Attack rate for ill persons who ate salad =
$$\frac{63}{115}$$
 X 100 = 54.8%

Attack rate for ill persons who did not eat salad =
$$\frac{10}{20}$$
 X 100 = 50%

$$RR = \frac{\text{Attack rate for ill persons who ate salad}}{\text{Attack rate for ill persons who did not eat salad}} = \frac{63/115}{10/20}$$

$$=$$
 $\frac{.548}{.50}$ $=$ 1.1

Interpretation: About 55% of the persons who became ill had eaten the salad served at the reception compared to 50% who became ill and had not eaten the salad. In other words, the risk of illness among persons who ate salad served at the reception was almost the same as the risk of illness among persons who did not eat the salad.

EATING DINNER ROLLS vs. BECOMING ILL

_	Ill	Not Ill	
Ate dinner rolls	57	40	97
Did not eat dinner rolls	15	23	38
•	72	63	135

Attack rate for ill persons who ate rolls =
$$\frac{57}{97}$$
 X 100 = 58.8%

Attack rate for ill persons who did not eat rolls =
$$\frac{15}{38}$$
 X 100 = 39.5%

$$=\frac{.588}{.395}=1.49$$

Interpretation: About 59% of the persons who became ill had eaten the dinner rolls served at the reception compared to 40% who became ill and had not eaten the dinner rolls. In other words, the risk of illness among persons who ate the dinner rolls served at the reception was 1.5 times higher than the risk of illness among persons who did not eat the dinner rolls.

EATING CHICKEN vs. BECOMING ILL

_	Ill	Not Ill	
Ate chicken	66	57	123
Did not eat chicken	6	6	12
	72	63	135

Attack rate for ill persons who ate chicken =
$$\frac{66}{123}$$
 X 100 = 53.7%

Attack rate for ill persons who did not eat chicken =
$$\frac{6}{12}$$
 X 100 = 50%

$$RR = \frac{\text{Attack rate for ill persons who ate chicken}}{\text{Attack rate for ill persons who did not eat chicken}} = \frac{66/123}{6/12}$$

$$=$$
 $\frac{.537}{.50}$ $=$ 1.1

Interpretation: About 54% of the persons who became ill had eaten the chicken served at the reception compared to 50% who became ill and had not eaten the chicken. In other words, the risk of illness among persons who ate chicken served at the reception was almost the same as the risk of illness among persons who did not eat the chicken.

EATING CAKE vs. BECOMING ILL

_	Ill	Not Ill	_
Ate cake	66	28	94
Did not eat cake	6	35	41
•	72	63	135

Attack rate for ill persons who ate cake =
$$\frac{66}{94}$$
 X 100 = 70.2%

Attack rate for ill persons who did not eat cake =
$$\frac{6}{41}$$
 X 100 = 14.6%

$$RR = \frac{\text{Attack rate for ill persons who ate cake}}{\text{Attack rate for ill persons who did not eat cake}} = \frac{66/94}{6/41}$$

$$=$$
 $\frac{.702}{.146}$ $=$ 4.8

Interpretation: About 66% of the persons who became ill had eaten the cake served at the reception compared to 24% who became ill and had not eaten the cake. In other words, the risk of illness among persons who ate cake served at the reception was almost 5 times higher than the risk of illness among persons who did not eat cake.

OVERALL INTERPRETATION: Based on the attack rates and relative risk ratios for each food item served at the reception, cake appears to be the suspect food item.

CALCULATIONS FOR CASE-CONTROL STUDIES

The 2 x 2 table may also be used to calculate the odds ratio for case-control studies.

The **ODDS RATIO** (**OR**) measures whether a specific exposure is associated with a certain disease. In other words, it is the ratio of the odds that the cases were exposed to the odds that the controls were exposed. Odds ratios related to exposure to specific food items can also be calculated.

NOTE: The relative risk ratio cannot be used to measure the association between exposure and illness for case-control studies because the total population exposed is not well-defined.

	Ill	Not Ill	
Exposed	a	b	$(\mathbf{a} + \mathbf{b})$
Not Exposed	c	d	$(\mathbf{c} + \mathbf{d})$
	$(\mathbf{a} + \mathbf{c})$	$(\mathbf{b} + \mathbf{d})$	a+b+c+d

Odds that the cases were exposed =
$$\frac{a}{b}$$

Odds that the controls were exposed
$$= \frac{c}{d}$$

$$OR = \frac{Odds \text{ that the cases were exposed}}{Odds \text{ that the controls were exposed}} = \frac{a/b}{c/d} = \frac{ad}{bc}$$

Interpretation of Odds Ratio (OR) †:

 $\mathbf{OR} = \mathbf{1}$: The odds of exposure among cases is the same as the odds of exposure among controls.

OR>1: The odds of exposure among cases is higher than the odds of exposure among controls.

OR<1: The odds of exposure among cases is lower than the odds of exposure among controls.

[†] "E. coli O157:H7 Infection in Michigan" Computer-based Case Study. <u>www.phppo.cdc.gov/phtn/casestudies/computerbased</u>.

EXAMPLE: Five persons reported eating at Restaurant X and becoming ill. After conducting a case-control study, the following numbers were obtained. Calculate the odds ratio and interpret the results. Also calculate the odds of becoming ill from eating a beef dish served at Restaurant X.

ODDS OF BECOMING A CASE FROM EATING AT RESTAURANT X

_	Case	Control	
Ate at Restaurant X	25	30	55
Did not eat at Restaurant X	10	40	50
	35	70	105

$$OR = \frac{Odds \text{ that the cases ate at Restaurant}}{Odds \text{ that the controls ate at}} = \frac{25/30}{10/40} = \frac{25(40)}{30(10)} = \frac{1000}{300}$$
Restaurant

Interpretation: The odds of exposure to Restaurant X was 3 times higher among cases than controls. It can be concluded that eating at Restaurant X may have contributed to illness.

ODDS OF BECOMING A CASE FROM EATING BEEF AT RESTAURANT X

_	Case	Control	
Ate beef at Restaurant X	30	13	43
Did not eat beef at Restaurant X	3	9	12
	33	22	_ 55

OR =
$$\frac{\text{Odds that the cases ate beef}}{\text{Odds that the controls ate beef}} = \frac{30/13}{3/9} = \frac{30(9)}{13(3)} = \frac{270}{39}$$

= 6.9

Interpretation: The odds of exposure to beef at Restaurant X was almost 7 times higher among cases than controls. It can be concluded that eating beef at Restaurant X may have contributed to illness.

SIGNIFICANCE TESTING OF THE RR AND THE OR

Tests of significance are calculated to determine if the association between exposure and illness occurred by chance alone. In other words, was the association observed between exposure and illness a random occurrence? The **95% confidence intervals** and the *p*-values may be calculated to determine the significance of the association between exposure and illness.

The 95% confidence intervals indicate how "confident" one can be that the RR or the OR observed actually lies within a range of numbers. In contrast, *p*-values represent the probability that the association observed between exposure and illness could have occurred by chance alone. Many statistical programs, like EpiInfoTM, readily calculate these values. Refer to these statistical programs or statistical books for more information.

APPENDIX F Final Epidemiology Report

F.1 Writing the Final Report

FORMAT FOR WRITING AN OUTBREAK REPORT

The following is a standard format of a written outbreak report. The format may be modified depending on the complexity of the outbreak.

INTRODUCTION BACKGROUND METHODS

Epidemiologic

Environmental

Laboratory and clinical

RESULTS

Epidemiologic

Environmental

Laboratory and clinical

DISCUSSION

RECOMMENDATIONS

ACKNOWLEDGEMENTS

SUPPORTING DOCUMENTS

The **INTRODUCTION** should include the following information:

- Who first reported the outbreak?
- When was the outbreak reported?
- What steps were taken to determine that an outbreak had occurred?
- What entities were involved with initiating the outbreak investigation?

The **BACKGROUND** should include the following information:

- What are the circumstances surrounding the outbreak?
- Where did the outbreak occur?
- What preliminary information was known?
 - o Demographics of the affected group
 - o Number of persons exposed
 - Number of persons ill
 - o Severity and clinical picture of ill persons

The **METHODS** section should include epidemiologic, environmental, and laboratory or clinical information:

- Epidemiologic
 - o What was the case definition?
 - o What investigation tools were used to collect or organize the information?
 - Line list
 - Epidemic curves
 - Maps
 - Chart reviews
 - Communication with health care providers
 - Questionnaire

- o If a questionnaire was administered, how was it administered?
 - Self-administered?
 - By phone?
 - In person?
 - Electronically?
- o What type of study was conducted?
- o What statistical analyses were conducted?
- o What hypotheses were generated?
- o What prevention and control measures were implemented?
- o What entities were involved?
- o What specific tasks were conducted?
- Environmental
 - o What kind of environmental investigation was conducted?
 - o Was a HACCP investigation performed?
 - o Were there any tracebacks?
 - o What was the physical layout of the outbreak?
- Laboratory and clinical
 - o Were stool samples collected for testing?
 - o Were food samples collected for testing?
 - o What tests were conducted?
 - o Where was testing performed?

The **RESULTS** section should also include epidemiologic, environmental, and laboratory or clinical information:

- Epidemiologic
 - o Total number of persons exposed (cohort study)
 - o For all individuals (cohort or case-control study)
 - Number and percentage of persons by age group
 - Number and percentage of persons by sex
 - Number and percentage of other demographics collected
 - Number and percentage of ill ("cases") and not-ill ("controls")
 - o For ill persons or cases (cohort or case-control study)
 - Number and percentage of each symptom experienced
 - Number of samples collected
 - Number and percentage of positive results
 - Number and percentage of persons hospitalized
 - Number and percentage of medical visits
 - Incubation period (median and range)
 - Recovery period (median and range)
 - o Overall attack rates and food-specific attack rates (cohort study)
 - Measures of association
 - Relative risk (cohort study)
 - Odds ratio (case-control study)
 - o Any additional results
- Environmental

- What were the results of the investigation or inspection, including any HACCP assessment?
- o What were the results of any food tracebacks, if done?
- What were potential environmental factors that may have contributed to the outbreak?
- Laboratory and clinical
 - o What were the results of the human samples submitted?
 - o What were the results of any food samples tested?

The **DISCUSSION** section should make interpretations of all the information collected during the outbreak investigation.

- Taking into account all the information collected, what can be concluded about the outbreak?
- Did the results from the epidemiologic investigation, environmental investigation, and laboratory testing support the hypotheses generated?
- Were there any important or interesting outcomes or findings?

The **RECOMMENDATIONS** section should provide educational information to aid others in outbreak investigations.

- What can be learned from this outbreak?
- Were the prevention and control measures implemented successful?
- What measures would prevent future occurrences?

The **ACKNOWLEDGEMENTS** section should recognize personnel who assisted in the outbreak investigation.

The **SUPPORTING DOCUMENTS** section should include any relevant information. Important documentation includes the following:

- Copy of the questionnaire or survey tool used
- Tables, epidemic curves, or maps
- Inspection reports

APPENDIX G Foodborne Illness and Etiology Tables

- **G.1** Etiologic Agents to Consider for Various Manifestations of Foodborne Illness
- **G.2** Foodborne Illnesses
- **G.3** Guidelines for Laboratory Confirmation of a Foodborne Disease Outbreak

G.1 Etiologic Agents to Consider for Various Manifestations of Foodborne Illness
Taken from Centers for Disease Control and Prevention. "Diagnosis and Management of Foodborne Illnesses: A Primer for Physicians and other Health Care Professionals." <i>MMWR</i> 2004:53 (No. RR-4).

TABLE 1. Etiologic agents to	consider for various	manifestations	of foodborne illness

Clinical presentation	Potential food-related agents to consider
Gastroenteritis (vomiting as primary symptom; fever and/or diarrhea also may be present)	Viral gastroenteritis, most commonly rotavirus in an infant or norovirus and other caliciviruses in an older child or adult; or food poisoning due to preformed toxins (eg, vomitoxin, <i>Staphylococcus aureus</i> toxin, <i>Bacillus cereus</i> toxin) and heavy metals.
Noninflammatory diarrhea (acute watery diarrhea without fever/dysentery; some patients may present with fever)*	Can be caused by virtually all enteric pathogens (bacterial, viral, parasitic) but is a classic symptom of Enterotoxigenic <i>Escherichia coli Giardia Vibrio cholerae</i> Enteric viruses (astroviruses, noroviruses and other caliciviruses, enteric adenovirus, rotavirus) <i>Cryptosporidium Cyclospora cayetanensis</i>
Inflammatory diarrhea (invasive gastroenteritis; grossly bloody stool and fever may be present) [†]	Shigella species Campylobacter species Salmonella species Enteroinvasive E. coli Enterohemorrhagic E. coli E. coli O157:H7 Vibrio parahaemolyticus Yersinia enterocolitica Entamoeba histolytica
Persistent diarrhea (lasting ≥14 days)	Prolonged illness should prompt examination for parasites, particularly in travelers to mountainous or other areas where untreated water is consumed. Consider <i>Cyclospora cayetanensis</i> , <i>Cryptosporidium</i> , <i>Entamoeba histolytica</i> , and <i>Giardia lamblia</i> .
Neurologic manifestations (eg, paresthesias, respiratory depression, bronchospasm, cranial nerve palsies)	Botulism (Clostridium botulinum toxin) Organophosphate pesticides Thallium poisoning Scombroid fish poisoning (histamine, saurine) Ciguatera fish poisoning (ciguatoxin) Tetradon fish poisoning (tetradotoxin) Neurotoxic shellfish poisoning (brevitoxin) Paralytic shellfish poisoning (saxitoxin) Amnesic shellfish poisoning (domoic acid) Mushroom poisoning Guillain-Barré syndrome (associated with infectious diarrhea due to Campylobacter jejuni)
Systemic illness (eg, fever, weakness, arthritis, jaundice)	Listeria monocytogenes Brucella species Trichinella spiralis Toxoplasma gondii Vibrio vulnificus Hepatitis A and E viruses Salmonella Typhi and Salmonella Paratyphi Amebic liver abscess

^{*} Noninflammatory diarrhea is characterized by mucosal hypersecretion or decreased absorption without mucosal destruction and generally involves the small intestine. Some affected patients may be dehydrated because of severe watery diarrhea and may appear seriously ill. This is more common in the young and the elderly. Most patients experience minimal dehydration and appear mildly ill with scant physical findings. Illness typically occurs with abrupt onset and brief duration. Fever and systemic symptoms usually are absent (except for symptoms related directly to intestinal fluid loss).

[†]Inflammatory diarrhea is characterized by mucosal invasion with resulting inflammation and is caused by invasive or cytotoxigenic microbial pathogens. The diarrheal illness usually involves the large intestine and may be associated with fever, abdominal pain and tenderness, headache, nausea, vomiting, malaise, and myalgia. Stools may be bloody and may contain many fecal leukocytes.

G.2	Foodborne Illnesses
	and Prevention. "Diagnosis and Management of Foodborne Illnessess ner Health Care Professionals." <i>MMWR</i> 2004:53 (No. RR-4).

Foodborne IIII	-	ciiai)	Domest's C			
Etiology	Incubation Period	Signs and Symptoms	Duration of Illness	Associated Foods	Laboratory Testing	Treatment
Bacillus anthracis	2 days to weeks	Nausea, vomiting, malaise, bloody diarrhea, acute abdominal pain.	Weeks	Insufficiently cooked contaminated meat.	Blood.	Penicillin is first choice for naturally acquired gastrointes- tinal anthrax. Ciprofloxacin is second option.
Bacillus cereus (preformed enterotoxin)	1–6 hrs	Sudden onset of severe nausea and vomiting. Diarrhea may be present.	24 hrs	Improperly refrigerated cooked or fried rice, meats.	Normally a clinical diagnosis. Clinical laboratories do not routinely identify this organism. If indicated, send stool and food specimens to reference laboratory for culture and toxin identification.	Supportive care.
Bacillus cereus (diarrheal toxin)	10–16 hours	Abdominal cramps, watery diarrhea, nausea.	24–48 hours	Meats, stews, gravies, vanilla sauce.	Testing not necessary, self- limiting (consider testing food and stool for toxin in outbreaks).	Supportive care.
Brucella abortus, B. melitensis, and B. suis	7–21 days	Fever, chills, sweating, weakness, headache, muscle and joint pain, diarrhea, bloody stools during acute phase.	Weeks	Raw milk, goat cheese made from unpasteurized milk, contaminated meats.	Blood culture and positive serology.	Acute: Rifampin and doxycycline daily for ≥6 weeks Infections with complications require combination therapy with rifampin, tetracycline, and an aminoglycoside.
Campylobacter jejuni	2–5 days	Diarrhea, cramps, fever, and vomiting; diarrhea may be bloody.	2–10 days	Raw and undercooked poultry, unpasturized milk, contaminated water.	Routine stool culture; Campylobacter requires special media and incubation at 42°C to grow.	Supportive care. For severe cases, antibiotics such as erythromycin and quinolones may be indicated early in the diarrheal disease. Guillain-Barré syndrome can be a sequela.
Clostridium botulinum— children and adults (preformed toxin)	12–72 hrs	Vomiting, diarrhea, blurred vision, diplopia, dysphagia, and descending muscle weakness.	Variable (from days to months). Can be compli- cated by respiratory failure and death.	Home-canned foods with a low acid content, improperly canned commercial foods, home-canned or fermented fish, herbinfused oils, baked potatoes in aluminium foil, cheese sauce, bottled garlic, foods held warm for extended periods of time (eg, in a warm oven).	Stool, serum, and food can be tested for toxin. Stool and food can also be cultured for the organism. These tests can be performed at some state health department laboratories and CDC.	Supportive care. Botulinum antitoxin is helpful if given early in the course of the illness. Contact the state health department. The 24-hour number for state health departments to call is (770) 488-7100.
Clostridium botulinum—infants	3–30 days	In infants <12 months, lethargy, weakness, poor feeding, constipation, hypotonia, poor head control, poor gag and sucking reflex.	Variable	Honey, home-canned vegetables and fruits, corn syrup.	Stool, serum, and food can be tested for toxin. Stool and food can also be cultured for the organism. These tests can be performed at some state health department laboratories and CDC.	Supportive care. Botulism immune globulin can be obtained from the Infant Botulism Prevention Program, Health and Human Services, California (510-540-2646). Botulinum antitoxin is generally not recommended for infants.
Clostridium perfringens toxin	8–16 hrs	Watery diarrhea, nausea, abdominal cramps; fever is rare.	24–48 hrs	Meats, poultry, gravy, dried or precooked foods, time- and/or temperature-abused food.	Stools can be tested for enterotoxin and cultured for organism. Because <i>Clostridium perfringens</i> can normally be found in stool, quantitative cultures must be done.	Supportive care. Antibiotics no indicated.
Enterohemorrhagic E. coli (EHEC) including E. coli O157:H7 and other Shiga toxin-producing E. coli (STEC)	1–8 days	Severe diarrhea that is often bloody, abdominal pain and vomiting. Usually, little or no fever is present. More common in children <4 years.	5–10 days	Undercooked beef especially hamburger, unpasteurized milk and juice, raw fruits and vegetables (eg. sprouts), salami (rarely), and contaminated water.	Stool culture; <i>E. coli</i> O157:H7 requires special media to grow. If <i>E. coli</i> O157:H7 is suspected, specific testing must be requested. Shiga toxin testing may be done using commercial kits; positive isolates should be forwarded to public health laboratories for confirmation and serotyping.	Supportive care, monitor renal function, hemoglobin, and platelets closely. <i>E. coli</i> O157:H7 infection is also associated with hemolytic uremic syndrome (HUS), which can cause lifelong complications. Studies indicate that antibiotics may promote the development of HUS.

	Incubation		Duration of			
Etiology	Period	Signs and Symptoms	Illness	Associated Foods	Laboratory Testing	Treatment
Enterotoxigenic E. coli (ETEC)	1–3 days	Watery diarrhea, abdominal cramps, some vomiting.	3 to >7 days	Water or food contaminated with human feces.	Stool culture. ETEC requires special laboratory techniques for identification. If suspected, must request specific testing.	Supportive care. Antibiotics are rarely needed except in severe cases. Recommended antibiotics include TMP-SMX and quinolones.
Listeria monocytogenes	9–48 hrs for gastrointestinal symptoms, 2–6 weeks for invasive disease	Fever, muscle aches, and nausea or diarrhea. Pregnant women may have mild flu-like illness, and infection can lead to premature delivery or stillbirth. Elderly or immunocompromised patients may have bacteremia or meningitis.	Variable	Fresh soft cheeses, unpasteurized milk, inadequately pasteur- ized milk, ready-to-eat deli meats, hot dogs.	Blood or cerebrospinal fluid cultures. Asymptomatic fecal carriage occurs; therefore, stool culture usually not helpful. Antibody to listerolysin O may be helpful to identify outbreak retrospectively.	Supportive care and antibiotics; Intravenous ampicillin, penicillin, or TMP-SMX are recommended for invasive disease.
	At birth and infancy	Infants infected from mother at risk for sepsis or meningitis.				
Salmonella spp.	1–3 days	Diarrhea, fever, abdominal cramps, vomiting. S. Typhi and S. Paratyphi produce typhoid with insidious onset characterized by fever, headache, constipation, malaise, chills, and myalgia; diarrhea is uncommon, and vomiting is not usually severe.	4–7 days	Contaminated eggs, poultry, unpasteurized milk or juice, cheese, contaminated raw fruits and vegetables (alfalfa sprouts, melons). S. Typhi epidemics are often related to fecal contamination of water supplies or street-vended foods.	Routine stool cultures.	Supportive care. Other than for S. Typhi and S. Paratyphi, antibiotics are not indicated unless there is extra-intestinal spread, or the risk of extra-intestinal spread, or the infection. Consider ampicillin, gentamicin, TMP-SMX, or quinolones if indicated. A vaccine exists for S. Typhi.
Shigella spp.	24–48 hrs	Abdominal cramps, fever, and diarrhea. Stools may contain blood and mucus.	4–7 days	Food or water contaminated with human fecal material. Usually person-to-person spread, fecal-oral transmission. Ready-to-eat foods touched by infected food workers, eg, raw vegetables, salads, sandwiches.	Routine stool cultures.	Supportive care. TMP-SMX recommended in the US if organism is susceptible; nalidixic acid or other quinolones may be indicated if organism is resistant, especially in developing countries.
Staphylococcus aureus (preformed enterotoxin)	1–6 hrs	Sudden onset of severe nausea and vomiting. Abdominal cramps. Diarrhea and fever may be present.	24–48 hrs	Unrefrigerated or improperly refrigerated meats, potato and egg salads, cream pastries.	Normally a clinical diagnosis. Stool, vomitus, and food can be tested for toxin and cultured if indicated.	Supportive care.
Vibrio cholerae (toxin)	24–72 hrs	Profuse watery diarrhea and vomiting, which can lead to severe dehydration and death within hours.	3–7 days. Causes life- threatening dehydra- tion.	Contaminated water, fish, shellfish, street- vended food typically from Latin America or Asia.	Stool culture; Vibrio cholerae requires special media to grow. If V. cholerae is suspected, must request specific testing.	Supportive care with aggressive oral and intravenous rehydration. In cases of confirmed cholera, tetracycline or doxycycline is recommended for adults, and TMP-SMX for children (<8 years).
Vibrio para- haemolyticus	2–48 hrs	Watery diarrhea, abdominal cramps, nausea, vomiting.	2–5 days	Undercooked or raw seafood, such as fish, shellfish.	Stool cultures. <i>Vibrio</i> parahaemolyticus requires special media to grow. If <i>V. parahaemolyticus</i> is suspected, must request specific testing.	Supportive care. Antibiotics are recommended in severe cases: tetracycline, doxycycline, gentamicin, and cefotaxime.
Vibrio vulnificus	1–7 days	Vomiting, diarrhea, abdominal pain, bacteremia, and wound infections. More common in the immunocompromised, or in patients with chronic liver disease (presenting with bullous skin lesions). Can be fatal in patients with liver disease and the immunocompromised.	2–8 days	Undercooked or raw shellfish, especially oysters, other contaminated seafood, and open wounds exposed to sea water.	Stool, wound, or blood cultures. Vibrio vulnificus requires special media to grow. If V. vulnificus is suspected, must request specific testing.	Supportive care and antibiotics; tetracycline, doxycycline, and ceftazidime are recommended.

Foodborne IIIr	nesses (Bacte	erial) (<i>Continued</i>)				
Etiology	Incubation Period	Signs and Symptoms	Duration of Illness	Associated Foods	Laboratory Testing	Treatment
Yersinia enterocolytica and Y. pseudotuber- culosis	24–48 hrs	Appendicitis-like symptoms (diarrhea and vomiting, fever, and abdominal pain) occur primarily in older children and young adults. May have a scarlitiniform rash with <i>Y. pseudotuberculosis</i> .	1–3 weeks, usually self- limiting	Undercooked pork, unpasteurized milk, tofu, contaminated water. Infection has occurred in infants whose caregivers handled chitterlings.	Stool, vomitus, or blood	Supportive care. If septicemia or other invasive disease occurs, antibiotic therapy with gentamicin or cefotaxime (doxycycline and ciprofloxacin also effective).
Foodborne IIII	Incubation		Duration of			
Etiology	Period	Signs and Symptoms	Illness	Associated Foods	Laboratory Testing	Treatment
Hepatitis A	28 days average (15–50 days)	Diarrhea, dark urine, jaundice, and flu-like symptoms, i.e., fever, headache, nausea, and abdominal pain.	Variable, 2 weeks – 3 months	Shellfish harvested from contaminated waters, raw produce, contaminated drinking water, uncooked foods and cooked foods that are not reheated after contact with infected food handler.	Increase in ALT, bilirubin. Positive IgM and anti- hepatitis A antibodies.	Supportive care. Prevention with immunization.
Noroviruses (and other caliciviruses)	12–48 hrs	Nausea, vomiting, abdominal cramping, diarrhea, fever, myalgia, and some headache. Diarrhea is more prevalent in adults and vomiting is more prevalent in children.	12–60 hrs	Shellfish, fecally contaminated foods, ready-to-eat foods touched by infected food workers (salads, sandwiches, ice, cookies, fruit).	Routine RT-PCR and EM on fresh unpreserved stool samples. Clinical diagnosis, negative bacterial cultures. Stool is negative for WBCs.	Supportive care such as rehydration. Good hygiene.
Rotavirus	1–3 days	Vomiting, watery diarrhea, low-grade fever. Temporary lactose intolerance may occur. Infants and children, elderly, and immunocompromised are especially vulnerable.	4–8 days	Fecally contaminated foods. Ready-to-eat foods touched by infected food workers (salads, fruits).	Identification of virus in stool via immunoassay.	Supportive care. Severe diarrhea may require fluid and electrolyte replacement.
Other viral agents (astroviruses, adenoviruses, parvoviruses)	10–70 hrs	Nausea, vomiting, diarrhea, malaise, abdominal pain, headache, fever.	2–9 days	Fecally contaminated foods. Ready-to-eat foods touched by infected food workers. Some shellfish.	Identification of the virus in early acute stool samples. Serology. Commercial ELISA kits are now available for adenoviruses and astroviruses.	Supportive care, usually mild, self-limiting. Good hygiene.
Foodborne IIII	nesses (Paras	sitic)				
Etiology	Incubation Period	Signs and Symptoms	Duration of Illness	Associated Foods	Laboratory Testing	Treatment
Angiostrongylus cantonensis	1 week to ≥1 month	Severe headaches, nausea, vomiting, neck stiffness, paresthesias, hyperesthesias, seizures, and other neurologic abnormalities.	Several weeks to several months	Raw or undercooked intermediate hosts (eg, snails or slugs), infected paratenic (transport) hosts (eg, crabs, fresh water shrimp), fresh produce contaminated with intermediate or transport hosts.	Examination of CSF for elevated pressure, protein, leukocytes, and eosinophils; serologic testing using ELISA to detect antibodies to Angiostrongylus cantonensis.	Supportive care. Repeat lumbar punctures and use of corticosteroid therapy may be used for more severely ill patients.
Cryptosporidium	2–10 days	Diarrhea (usually watery), stomach cramps, upset stomach, slight fever.	May be remitting and relapsing over weeks to months	Any uncooked food or food contaminated by an ill food handler after cooking, drinking water.	Request specific examination of the stool for <i>Cryptosporidium</i> . May need to examine water or food.	Supportive care, self-limited. If severe consider paromomycin for 7 days. For children aged 1–11 years, consider nitazoxanide for 3 days.
Cyclospora cayetanensis	1–14 days, usually at least 1 week	Diarrhea (usually watery), loss of appetite, substantial loss of weight, stomach cramps, nausea, vomiting, fatigue.	May be remitting and relapsing over weeks to months	Various types of fresh produce (imported berries, lettuce).	Request specific examination of the stool for <i>Cyclospora</i> . May need to examine water or food.	TMP-SMX for 7 days.

Foodborne IIII	nesses (Para	sitic) (<i>Continued</i>)				
Etiology	Incubation Period	Signs and Symptoms	Duration of Illness	Associated Foods	Laboratory Testing	Treatment
Entamoeba histolytica	2–3 days to 1–4 weeks	Diarrhea (often bloody), frequent bowel move- ments, lower abdominal pain.	May be protracted (several weeks to several months)	Any uncooked food or food contaminated by an ill food handler after cooking, drinking water.	Examination of stool for cysts and parasites—may need at least 3 samples. Serology for long-term infections.	Metronidazole and a luminal agent (iodoquinol or paromomycin).
Giardia lamblia	1–2 weeks	Diarrhea, stomach cramps, gas.	Days to weeks	Any uncooked food or food contaminated by an ill food handler after cooking, drinking water.	Examination of stool for ova and parasites — may need at least 3 samples.	Metronidazole.
Toxoplasma gondii	5–23 days	Generally asymptomatic, 20% may develop cervical lymphadenopathy and/or a flu-like illness. In immunocompromised patients: central nervous system (CNS) disease, myocarditis, or pneumonitis is often seen.	Months	Accidental ingestion of contaminated substances (eg, soil contaminated with cat feces on fruits and vegetables), raw or partly cooked meat (especially pork, lamb, or venison).	Isolation of parasites from blood or other body fluids; observation of parasites in patient specimens via microscopy or histology. Detection of organisms is rare; serology (reference laboratory needed) can be a useful adjunct in diagnosing toxoplasmosis. However, IgM antibodies may persist for 6–18 months and thus may not necessarily indicate recent infection. PCR of bodily fluids. For congenital infection: isolation of <i>T. gondii</i> from placenta, umbilical cord, or infant blood. PCR of white blood cells, CSF, or amniotic fluid, or IgM and IgA serology, performed by a reference laboratory.	Asymptomatic healthy, but infected, persons do not require treatment. Spiramycin or pyrimethamine plus sulfadiazine may be used for pregnant women. Pyrimethamine plus sulfadiazine may be used for immunocompromised persons, in specific cases. Pyrimethamine plus sulfadiazine (with or without steroids) may be given for ocular disease when indicated Folinic acid is given with pyrimethamine plus sulfadiazine to counteract bone marrow suppression.
Toxoplasma gondii (congenital infection)	In infants at birth	Treatment of the mother may reduce severity and/ or incidence of congenital infection. Most infected infants have few symptoms at birth. Later, they will generally develop signs of congenital toxoplasmosis (mental retardation, severely impaired eyesight, cerebral palsy, seizures), unless the infection is treated.	Months	Passed from mother (who acquired acute infection during pregnancy) to child.		
Trichinella spiralis	1–2 days for initial symptoms; others begin 2–8 weeks after infection	Acute: nausea, diarrhea, vomiting, fatigue, fever, abdominal discomfort followed by muscle soreness, weakness, and occasional cardiac and neurologic complications.	Months	Raw or undercooked contaminated meat, usually pork or wild game meat (eg, bear or moose).	Positive serology or demonstration of larvae via muscle biopsy. Increase in eosinophils.	Supportive care plus mebendazole or albendazole.

	Incubation		Duration of			
Etiology	Period	Signs and Symptoms	Illness	Associated Foods	Laboratory Testing	Treatment
Antimony	5 min – 8 hrs. usually <1 hr	Vomiting, metallic taste.	Usually self-limited	Metallic container.	Identification of metal in beverage or food.	Supportive care.
Arsenic	Few hrs	Vomiting, colic, diarrhea.	Several days	Contaminated food.	Urine. May cause eosinophilia.	Gastric lavage, BAL (dimercaprol).
Cadmium	5 min – 8 hrs. usually <1 hr	Nausea, vomiting, myalgia, increase in salivation, stomach pain.	Usually self-limited	Seafood, oysters, clams, lobster, grains, peanuts.	Identification of metal in food.	Supportive care.
Ciguatera fish poisoning (ciguatera toxin)	2–6 hrs	GI: abdominal pain, nausea, vomiting, diarrhea.	Days to weeks to months	A variety of large reef fish. Grouper, red snapper, amberjack, and barracuda (most	Radioassay for toxin in fish or a consistent history.	Supportive care, IV mannitol. Children more vulnerable.
	3 hrs	Neurologic: paresthesias, reversal of hot or cold, pain, weakness.		common).		
	2–5 days	<u>Cardiovascular:</u> bradycardia, hypotension, increase in T wave abnormalities.				
Copper	5 min – 8 hrs. usually <1 hr	Nausea, vomiting, blue or green vomitus.	Usually self-limited	Metallic container.	Identification of metal in beverage or food.	Supportive care.
Mercury	1 week or longer	Numbness, weakness of legs, spastic paralysis, impaired vision, blindness, coma. Pregnant women and the developing fetus are especially vulnerable.	May be protracted	Fish exposed to organic mercury, grains treated with mercury fungicides.	Analysis of blood, hair.	Supportive care.
Mushroom toxins, short-acting (museinol, muscarine, psilocybin, coprius artemetaris, ibotenic acid)	<2 hrs	Vomiting, diarrhea, confusion, visual disturbance, salivation, diaphoresis, hallucinations, disulfiram-like reaction, confusion, visual disturbance.	Self-limited	Wild mushrooms (cooking may not destroy these toxins).	Typical syndrome and mushroom identified or demonstration of the toxin.	Supportive care.
Mushroom toxin, long-acting (amanitin)	4–8 hrs diarrhea; 24–48 hrs liver failure	Diarrhea, abdominal cramps, leading to hepatic and renal failure.	Often fatal	Mushrooms.	Typical syndrome and mushroom identified and/or demonstration of the toxin.	Supportive care, life- threatening, may need life support.
Nitrite poisoning	1–2 hrs	Nausea, vomiting, cyanosis, headache, dizziness, weakness, loss of consciousness, chocolate-brown colored blood.	Usually self-limited	Cured meats, any contaminated foods, spinach exposed to excessive nitrification.	Analysis of the food, blood.	Supportive care, methylene blue.
Pesticides (organophosphates or carbamates)	Few min to few hrs	Nausea, vomiting, abdominal cramps, diarrhea, headache, nervousness, blurred vision, twitching, convulsions, salivation and meiosis.	Usually self-limited	Any contaminated food.	Analysis of the food, blood.	Atropine; 2-PAM (Pralidoxime) is used when atropine is not able to control symptoms and is rarely necessary in carbamate poisoning.
Puffer fish (tetrodotoxin)	<30 min	Parasthesias, vomiting, diarrhea, abdominal pain, ascending paralysis, respiratory failure.	Death usually in 4–6 hours	Puffer fish.	Detection of tetrodotoxin in fish.	Life-threatening, may need respiratory support.
Scombroid (histamine)	1 min – 3 hrs	Flushing, rash, burning sensation of skin, mouth and throat, dizziness, uriticaria, parasthesias.	3–6 hrs	Fish: bluefin, tuna, skipjack, mackerel, marlin, escolar, and mahi mahi.	Demonstration of histamine in food or clinical diagnosis.	Supportive care, antihistamines.

Foodborne Illnesses (Noninfectious) (Continued)

Etiology	Incubation Period	Signs and Symptoms	Duration of Illness	Associated Foods	Laboratory Testing	Treatment
Shellfish toxins (diarrheic, neurotoxic, amnesic)	Diarrheic shellfish poisoning (DSP) — 30 min to 2 hrs	Nausea, vomiting, diarrhea, and abdominal pain accompanied by chills, headache, and fever.	Hrs to 2–3 days	A variety of shellfish, primarily mussels, oysters, scallops, and shellfish from the Florida coast and the Gulf of Mexico.	Detection of the toxin in shellfish; high-pressure liquid chromatography.	Supportive care, generally self- limiting. Elderly are especially sensitive to ASP.
	Neurotoxic shellfish poisoning (NSP) — few min to hours	Tingling and numbness of lips, tongue, and throat, muscular aches, dizziness, reversal of the sensations of hot and cold, diarrhea, and vomiting.		WEALCO.		
	Amnesic shellfish poisoning (ASP) — 24–48 hrs	Vomiting, diarrhea, abdominal pain and neurologic problems such as confusion, memory loss, disorientation, seizure, coma.				
Shellfish toxins (paralytic shellfish poisoning)	30 min – 3 hrs	Diarrhea, nausea, vomiting leading to parasthesias of mouth, lips, weakness, dysphasia, dysphonia, respiratory paralysis.	Days	Scallops, mussels, clams, cockles.	Detection of toxin in food or water where fish are located; high-pressure liquid chromatography.	Life-threatening, may need respiratory support.
Sodium fluoride	Few min to 2 hrs	Salty or soapy taste, numbness of mouth, vomiting, diarrhea, dilated pupils, spasms, pallor, shock, collapse.	Usually self-limited	Dry foods (eg, dry milk, flour, baking powder, cake mixes) contami- nated with sodium fluoride—containing insecticides and rodenticides.	Testing of vomitus or gastric washings. Analysis of the food.	Supportive care.
Thallium	Few hrs	Nausea, vomiting, diarrhea, painful parathesias, motor polyneuropathy, hair loss.	Several days	Contaminated food.	Urine, hair.	Supportive care.
Tin	5 min – 8 hrs. usually <1 hr	Nausea, vomiting, diarrhea.	Usually self-limited	Metallic container.	Analysis of the food.	Supportive care.
Vomitoxin	Few min to 3 hrs	Nausea, headache, abdominal pain, vomiting.	Usually self-limited	Grains such as wheat, corn, barley.	Analysis of the food.	Supportive care.
Zinc	Few hrs	Stomach cramps, nausea, vomiting, diarrhea, myalgias.	Usually self-limited	Metallic container.	Analysis of the food, blood and feces, saliva or urine.	Supportive care.

G.3	Guidelines for Laboratory Confirmation of a
	Foodborne Disease Outbreak

Taken from Centers for Disease Control and Prevention. "Diagnosis and Management of Foodborne Illnesses: A Primer for Physicians and other Health Care Professionals." *MMWR* 2001:50 (No. RR-2).

Etiologic agent	Incubation period	Clinical syndrome	Confirmation
Bacterial 1. Bacillus cereus a. Vomiting toxin	1–6 hrs	Vomiting; some patients with	Isolation of organism from stool of
· ·		diarrhea; fever uncommon	two or more ill persons and not from stool of control patients OR
			Isolation of 10 ⁵ organisms/g from epidemiologically implicated food, provided specimen is properly handled
b. Diarrheal toxin	6–24 hrs	Diarrhea, abdominal cramps, and vomiting in some patients; fever uncommon	Isolation of organism from stool of two or more ill persons and not from stool of control patients OR
			Isolation of 10 ⁵ organisms/g from epidemiologically implicated food, provided specimen is properly handled
2. Brucella	Several days to several mos; usually >30 days	Weakness, fever, headache, sweats, chills, arthralgia, weight loss, splenomegaly	Two or more ill persons and isolation of organism in culture of blood or bone marrow; greater than fourfold increase in standard agglutination titer (SAT) over several wks, or single SAT 1:160 in person who has compatible clinical symptoms and history of exposure
3. Campylobacter jejuni/coli	2–10 days; usually 2–5 days	Diarrhea (often bloody), abdominal pain, fever	Isolation of organism from clinical specimens from two or more ill persons OR Isolation of organism from epidemiologically implicated food
			G.3
	Taker	from MMWR 2001; 50 (No. RR-2)	1 of 8

0 h 0 d a		
2 hrs–8 days; usually 12–48 hrs	Illness of variable severity; common symptoms are diplopia, blurred vision, and bulbar weakness; paralysis, which is usually descending and bilateral, might progress rapidly	Detection of botulinal toxin in serum, stool, gastric contents, or implicated food OR Isolation or organism from stool or intestine
6–24 hrs	Diarrhea, abdominal cramps; vomiting and fever uncommon	Isolation of 10 ⁵ organisms/g from stool of two or more ill persons, provided specimen is properly handled. OR Demonstration of enterotoxin in the stool of two or more ill persons OR Isolation of 10 ⁵ organisms/g from epidemiologically implicated food, provided specimen is properly handled
1–10 days; usually 3–4 days	Diarrhea (often bloody), abdominal cramps (often severe), little or no fever	Isolation of <i>E. coli</i> O157:H7 or other Shiga-like toxin-producing <i>E. coli</i> from clinical specimen from two or more ill persons OR Isolation of <i>E. coli</i> O157:H7 or other Shiga-like toxin-producing <i>E. coli</i> from epidemiologically implicated food
6–48 hrs	Diarrhea, abdominal cramps, nausea; vomiting and fever less common	Isolation of organism of same serotype, demonstrated to produce heat-stable (ST) and/or heat-labile (LT) enterotoxin, from stool of two or more ill persons
Variable	Diarrhea, fever, abdominal cramps	Isolation of organism of same enteropathogenic serotype from stool of two or more ill persons
	1–10 days; usually 3–4 days 6–48 hrs	vision, and bulbar weakness; paralysis, which is usually descending and bilateral, might progress rapidly 6–24 hrs Diarrhea, abdominal cramps; vomiting and fever uncommon Diarrhea (often bloody), abdominal cramps (often severe), little or no fever Diarrhea, abdominal cramps, nausea; vomiting and fever less common

Etiologic agent	Incubation period	Clinical syndrome	Confirmation
d. Enteroinvasive (EIEC)	Variable	Diarrhea (might be bloody), fever, abdominal cramps	Isolation of same enteroinvasive serotype from stool of two or more ill persons
7. Listeria monocytogenes			
a. Invasive disease	2–6 wks	Meningitis, neonatal sepsis, fever	Isolation of organism from normally sterile site
b. Diarrheal disease	Unknown	Diarrhea, abdominal cramps, fever	Isolation of organism of same serotype from stool of two or more ill persons exposed to food that is epidemiologically implicated or from which organism of same serotype has been isolated
8. Nontyphoidal <i>Salmonella</i>	6 hrs-10 days; usually 6-48 hrs	Diarrhea, often with fever and abdominal cramps	Isolation of organism of same serotype from clinical specimens from two or more ill persons OR Isolation of organism from epidemiologically implicated food
9. <i>Salmonella</i> Typhi	3–60 days; usually 7–14 days	Fever, anorexia, malaise, headache, and myalgia; sometimes diarrhea or constipation	Isolation of organism from clinical specimens from two or more ill persons OR Isolation of organism from epidemiologically implicated food
10. <i>Shigella</i> spp.	12 hrs-6 days; usually 2-4 days	Diarrhea (often bloody), often accompanied by fever and abdominal cramps	Isolation of organism of same serotype from clinical specimens from two or more ill persons OR Isolation of organism from epidemiologically implicated food
			G.3
	Taken	from MMWR 2001; 50 (No. RR-2)	3 of 8

Etiologic agent	Incubation period	Clinical syndrome	Confirmation
11.Staphylococcus aureus	30 min-8 hrs; usually 2-4 hrs	Vomiting, diarrhea	Isolation of organism of same phage type from stool or vomitus of two or more ill persons OR
			Detection of enterotoxin in epidemiologically implicated food OR
			Isolation of 10 ⁵ organisms/g from epidemiologically implicated food, provided specimen is properly handled
12. <i>Streptococcus,</i> group A	1–4 days	Fever, pharyngitis, scarlet fever, upper respiratory infection	Isolation of organism of same M- or T-type from throats of two or more ill persons OR
			Isolation of organism of same M- or T-type from epidemiologically implicated food
13.Vibrio cholerae			
a.O1 or O139	1–5 days	Watery diarrhea, often accompanied by vomiting	Isolation of toxigenic organism from stool or vomitus of two or more ill persons OR
			Significant rise in vibriocidal, bacterial-agglutinating, or antitoxin antibodies in acute- and early convalescent-phase sera among persons not recently immunized OR
			Isolation of toxigenic organism from epidemiologically implicated food
b. non-O1 and non-O139	1–5 days	Watery diarrhea	Isolation of organism of same serotype from stool of two or more ill persons
		n from MMWR 2001; 50 (No. RR-2)	G.3

Etiologic agent	Incubation period	Clinical syndrome	Confirmation
14.Vibrio parahaemolyticus	4–30 hrs	Diarrhea	Isolation of Kanagawa-positive organism from stool of two or more ill persons OR Isolation of 10 ⁵ Kanagawa-positive organisms/g from epidemiologically implicated food, provided specimen is properly handled
15. Yersinia enterocolitica	1–10 days; usually 4–6 days	Diarrhea, abdominal pain (often severe)	Isolation of organism from clinical specimen from two or more ill persons OR Isolation of pathogenic strain of organism from epidemiologically implicated food
Chemical			
1. Marine toxins a. Ciguatoxin	1–48 hrs; usually 2–8 hrs	Usually gastrointestinal symptoms followed by neurologic symptoms (including paresthesia of lips, tongue, throat, or extremities) and reversal of hot and cold sensation	Demonstration of ciguatoxin in epidemiologically implicated fish OR Clinical syndrome among persons who have eaten a type of fish previously associated with ciguatera fish poisoning (e.g., snapper, grouper, or barracuda)
b. Scombroid toxin (histamine)	1 min–3 hrs; usually <1 hr	Flushing, dizziness, burning of mouth and throat, headache, gastrointestinal symptoms, urticaria, and generalized pruritis	Demonstration of histamine in epidemiologically implicated fish OR Clinical syndrome among persons who have eaten a type of fish previously associated with histamine fish poisoning (e.g., mahi-mahi or fish of order Scomboidei)
			G.3
	Take	n from MMWR 2001; 50 (No. RR-2)	5 of 8

Etiologic agent	Incubation period	Clinical syndrome	Confirmation
c. Paralytic or neurotoxic shellfish	30 min–3 hrs	Paresthesia of lips, mouth or face, and extremities; intestinal symptoms or weakness, including respiratory difficulty	Detection of toxin in epidemiologically implicated food OR Detection of large numbers of shellfish-poisoning-associated species of dinoflagellates in water from which epidemiologically
d. Puffer fish, tetrodotoxin	10 min–3 hrs; usually 10–45 min	Paresthesia of lips, tongue, face, or extremities, often following numbness; loss of proprioception or floating sensations	implicated mollusks are gathered Demonstration of tetrodotoxin in epidemiologically implicated fish OR Clinical syndrome among persons who have eaten puffer fish
2. Heavy metalsAntimonyCadmiumCopperIronTinZinc	5 min–8 hrs; usually <1 hr	Vomiting, often metallic taste	Demonstration of high concentration of metal in epidemiologically implicated food
3. Monosodium glutamate (MSG)	3 min–2 hrs; usually <1 hr	Burning sensation in chest, neck, abdomen, or extremities; sensation of lightness and pressure over face or heavy feeling in chest	Clinical syndrome among persons who have eaten food containing MSG (e.g., usually 1.5 g MSG)
 4. Mushroom toxins a. Shorter-acting toxins Muscimol Muscarine Psilocybin Coprinus artrementa Ibotenic acid 	2 hrs	Usually vomiting and diarrhea, other symptoms differ with toxin • Confusion, visual disturbance • Salivation, diaphoresis • Hallucinations • Disulfiram-like reaction • Confusion, visual disturbance	Clinical syndrome among persons who have eaten mushroom identified as toxic type OR Demonstration of toxin in epidemiologically implicated mushroom or food containing mushroom
			G.3
		Taken from MMWR 2001; 50 (No. RR-2)	6 of 8

Etiologic agent	Incubation period	Clinical syndrome	Confirmation
b. Longer-acting toxins (e.g., <i>Amanita</i> spp.)	6–24 hrs	Diarrhea and abdominal cramps for 24 hrs followed by hepatic and renal failure	Clinical syndrome among persons who have eaten mushroom identified as toxic type OR Demonstration of toxin in epidemiologically implicated mushroom or food containing mushrooms
Parasitic			
1. Cryptosporidium parvum	2–28 days; median: 7 days	Diarrhea, nausea, vomiting; fever	Demonstration of organism or antigen in stool or in small-bowel biopsy of two or more ill persons OR
			Demonstration of toxin in epidemiologically implicated food
Cyclospora cayetanensus	1–11 days; median: 7 days	Fatigue, protracted diarrhea, often relapsing	Demonstration of organism in stool of two or more ill persons
3. Giardia lamblia	3–25 days; median: 7 days	Diarrhea, gas, cramps, nausea, fatigue	Two or more ill persons and detection of antigen in stool or demonstration of organism in stool, duodenal contents, or small-bowel biopsy specimen
4. <i>Trichinella</i> spp.	1–2 days for intestinal phase; 2–4 wks for systemic phase	Fever, myalgia, periorbital edema, high eosinophil count	Two or more ill persons and positive serologic test or demonstration of larvae in muscle biopsy OR Demonstration of larvae in
			epidemiologically implicated meat
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Viral 1. Hepatitis A 15–50 days; median: 28 days Jaundice, dark urine, fatigue, anorexia, nausea Jaundice, dark urine, fatigue, anti-patigue, anti-patigue	Etiologic agent	Incubation period	Clinical syndrome	Confirmation
2. Norwalk family of viruses, small round-structured viruses (SRSV) 15-77 hrs; usually 24-48 hrs viruses had convalescent sera in most serum pairs OR Visualization of small, round-structured viruses that react with patient's convalescent sera by immune-electron microsopy (assays based on molecular diagnostics [e.g., polymerase-chain reaction, probes, or assays for antigen and antibodies from expressed antigen] are available in reference laboratories) Vomiting, cramps, diarrhea, headache in reference laboratories) Visualization of small, round-structured viruses that react with patient's convalescent sera but not acute sera — by immune-electron microsopy (assays based on molecular diagnostics [e.g., polymerase-chain reaction, probes, or assays for antigen and antibodies from expressed antigen] are available in reference laboratories)		15–50 days; median: 28 days		anti-hepatitis A virus in serum from two or more persons who consumed epidemiologically
3. Astrovirus, calicivirus, others 15–77 hrs; usually 24–48 hrs others Vomiting, cramps, diarrhea, headache Visualization of small, round-structured viruses that react with patient's convalescent sera but not acute sera — by immune-electron microsopy (assays based on molecular diagnostics [e.g., polymerase-chain reaction, probes, or assays for antigen and antibodies from expressed antigen] are available in reference laboratories) G.3	viruses, small round-structured	15–77 hrs; usually 24–48 hrs		More than fourfold rise in antibody titer to Norwalk virus or Norwalk-like virus in acute and convalescent sera in most serum pairs OR Visualization of small, round-structured viruses that react with patient's convalescent sera but not acute sera — by immune-electron microsopy (assays based on molecular diagnostics [e.g., polymerase-chain reaction, probes, or assays for antigen and antibodies from expressed antigen] are available
		15–77 hrs; usually 24–48 hrs		Visualization of small, round-structured viruses that react with patient's convalescent sera but not acute sera — by immune-electron microsopy (assays based on molecular diagnostics [e.g., polymerase-chain reaction, probes, or assays for antigen and antibodies from expressed antigen] are available in reference laboratories)
		Takas	n from MMWR 2001: 50 (No. P.P. 2)	

APPENDIX H Foodborne Illness Complaint Worksheet used by Food Inspectors

H.1 Foodborne Illness Investigation Form 910

FPCS -910 1/02

KANSAS DEPARTMENT OF HEALTH AND ENVIRONMENT

Foodborne Illness Investigation Report

Date Received:	Occurrence I	Date:	
Event Location:	City:	County:	
Establishment Food Prepared At:		ID#:	
How Many People Served:	Number Repo	orted Ill:	
Time Food Consumed:	Time of Onse	et (AM/PM):	
Symptoms:			
☐ Nausea ☐ Vomiting ☐ Diarrhea ☐	Bloody Diarrhea	☐ Abdominal Pain	☐ Dizziness
☐ Blurred Vision ☐ Headache ☐ Other: _			
Suspected Foods:			
Samples Taken?	Date Samples Sent T	o Lab:	
Name of EPI Contact:	Time:	Date: _	
Date HACCP Inspection Conducted:	Date F	IACCP Scheduled:	
Comments:			
Inspector:	Investigation	Date:	
Travel Time:	Total Investig	gation Time:	

APPENDIX I Infected Food Handlers

- I.1 Exclusion and Restriction Requirements for Infected Food handlers
- I.2 Removal of Exclusion and Restriction Requirements for Infected Food handlers

EXCLUSION AND RESTRICTION REQUIREMENTS FOR INFECTED FOODHANDLERS

Food handlers with specific diseases or health conditions should be excluded and restricted according to the following requirements. These requirements are based on the 1999 Kansas Food Code. Such

Health Status of Food handler	Facilities Serving Highly Susceptible Population [†] T	Facilities Not Serving Highly Susceptible Population
Diagnosed with illness due to <i>Salmonella</i> Typhi, <i>Shigella spp.</i> , Shiga toxin-producing <i>Escherchia coli</i> , or hepatitis A virus	EXCLUDE	EXCLUDE
Experiencing one of the following symptoms: diarrhea, fever, vomiting, jaundice, sore through with fever <i>OR</i> experiencing an uncovered lesion containing pus on the hands or wrists, exposed portions of arms, or other parts of the body	RESTRICT	RESTRICT
Experiencing one of the following symptoms and meets a high-risk condition [‡] : diarrhea, fever, vomiting, jaundice, sore throat with fever	EXCLUDE	RESTRICT
Asymptomatic but stools positive for <i>S</i> . Typhi, <i>Shigella spp</i> ., or Shiga toxin-producing E. coli	EXCLUDE	RESTRICT
Past illness from <i>Salmonella</i> Typhi within the last 3 months	EXCLUDE	NO RESTRICTIONS
Past illness from <i>Shigella spp</i> . or Shiga toxin-producing <i>Escherichia coli</i> within the last month	EXCLUDE	NO RESTRICTIONS
Onset of jaundice within the last 14 days	EXCLUDE	EXCLUDE
Onset of jaundice more than 14 days before	EXCLUDE	RESTRICT

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[†] A group of persons who are more likely than other populations to experience foodborne disease because they are immunocompromised or older adults and in a facility that provides health care or assisted living services, such as a hospital or nursing home; or preschool age children in a facility that provides custodial care, such as a day care center.

[‡] A food handler with high-risk conditions refers to a food handler who (1) prepared or consumed food that caused disease, lives with an ill person, or lives with a person associated with a disease outbreak. The diseases of concern include *S. typhi*, *Shigella spp.*, Shiga-toxin producing *E. coli*, or hepatitis A virus.

REMOVAL OF EXCLUSION AND RESTRICTION REQUIREMENTS FOR FOODHANDLERS

Infected food handlers who have been restricted or excluded may be removed from such requirements after the following criteria are met. These requirements are based on the 1999 Kansas Food Code.

Health Status of Food handler	Facilities Serving Highly Susceptible Population [†]	Facilities Not Serving Highly Susceptible Population
Diagnosed with illness due to Salmonella Typhi Diagnosed with illness due to Shigella spp. or Shiga toxin-producing Escherchia coli	 Approval from regulatory authority and Written medical documentation of 3 consecutive negative stools taken 1 month after onset, 48 hours after discontinuance of antibiotics, and 24 hours apart Approval from regulatory authority and Written medical documentation of 2 consecutive negative stools taken 48 hours after discontinuance of antibiotics and 24 hours apart 	 Approval from regulatory authority and Written medical documentation of 3 consecutive negative stools taken 1 month after onset, 48 hours after discontinuance of antibiotics, and 24 hours apart Approval from regulatory authority and Written medical documentation of 2 consecutive negative stools taken 48 hours after discontinuance of antibiotics and 24 hours apart
Diagnosed with illness due to hepatitis A virus	 Approval from regulatory authority and Written medical documentation that person is free of symptoms or has 2 blood tests demonstrating falling liver enzymes 	 Approval from regulatory authority and Written medical documentation that person is free of symptoms or has 2 blood tests demonstrating falling liver enzymes
Experiencing one of the following symptoms: diarrhea, fever, vomiting, jaundice, sore through with fever <i>OR</i> experiencing an uncovered lesion containing pus on the hands or wrists, exposed portions of arms, or other parts of the body	 Person is free of symptoms and no illnesses resulted and Written medical documentation that person is free of symptoms, free of infectious agent, or has noninfectious condition 	 Person is free of symptoms and no illnesses resulted and Written medical documentation that person is free of symptoms, free of infectious agent, or has noninfectious condition

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[†] A group of persons who are more likely than other populations to experience foodborne disease because they are immunocompromised or older adults and in a facility that provides health care or assisted living services, such as a hospital or nursing home; or preschool age children in a facility that provides custodial care, such as a day care center.

REMOVAL OF EXCLUSION AND RESTRICTION REQUIREMENTS FOR FOODHANDLERS (continued)

Health Status of Food handler	Facilities Serving Highly Susceptible Population [†]	Facilities Not Serving Highly Susceptible Population
Experiencing one of the following symptoms and meets a high-risk condition [‡] : diarrhea, fever, vomiting, jaundice, sore throat with fever	 Written medical documentation that person is free of infectious agent, is free of jaundice, or has noninfectious condition 	 Written medical documentation that person is free of infectious agent, is free of jaundice, or has noninfectious condition
Asymptomatic but stools positive for <i>S</i> . Typhi, <i>Shigella spp</i> ., or Shiga toxin-producing E. coli	 Written medical documentation that person is free of infectious agent 	 Written medical documentation that person is free of infectious agent
Past illness from Salmonella Typhi within the last three months	 Written medical documentation that person is free of infectious agent 	NA
Past illness from Shigella spp. Or Shiga toxin- producing Escherichia coli within the last month	 Written medical documentation that person is free of infectious agent 	NA
Onset of jaundice within the last 14 days	 No illnesses resulted and Written medical documentation that person is free of hepatitis A or has 2 blood tests demonstrating falling liver enzymes 	 No illnesses resulted and Written medical documentation that person is free of hepatitis A or has 2 blood tests demonstrating falling liver enzymes
Onset of jaundice more than 14 days before	 Until 2 weeks after onset of symptoms and Written medical documentation that person is free of hepatitis A or has 2 blood tests demonstrating falling liver enzymes 	 Until 2 weeks after onset of symptoms and Written medical documentation that person is free of hepatitis A or has 2 blood tests demonstrating falling liver enzymes

[†] A group of persons who are more likely than other populations to experience foodborne disease because they are immunocompromised or older adults and in a facility that provides health care or assisted living services, such as a hospital or nursing home; or preschool age children in a facility that provides custodial care, such as a day care center.

[‡] A food handler with high-risk conditions refers to a food handler who (1) prepared or consumed food that caused disease, lives with an ill person, or lives with a person associated with a disease outbreak. The diseases of concern include *S. typhi*, *Shigella spp.*, Shiga-toxin producing *E. coli*, or hepatitis A virus.

APPENDIX J Hazard Analysis Critical Control Points (HACCP)

J.1 Overview and HACCP Inspections

HAZARD ANALYSIS AND CRITICAL CONTROL POINT (HACCP)

Overview

Hazard Analysis and Critical Control Point, or HACCP (pronounced HAS-SIP), is a systematic, science-based approach of identifying, evaluating, and controlling food safety hazards[†]. Initially developed to keep food safe for astronauts within the space program, this approach was adopted by the Food and Drug Administration and the U.S. Department of Agriculture as a means of ensuring a safe food supply from harvest to consumption. Currently, the seafood industry, juice industry, and meat and poultry processing plants are required to follow a HACCP plan, or a written documentation of all food processing and handling procedures. A number of food companies in the U. S. have also adopted a HACCP plan in their manufacturing processes.

The following table lists the seven fundamental HACCP principles.

HACCP Principles[†]

- 1. Conduct a hazard analysis
- 2. Identify the critical control points (CCP)
- 3. Establish critical limits for each CCP
- 4. Establish monitoring procedures
- 5. Establish corrective actions
- 6. Establish recordkeeping procedures
- 7. Establish verification procedures

PRINCIPLE #1: Conduct a hazard analysis

A hazard analysis is the identification of (1) potential biological, chemical, or physical threats to food and (2) preventative measures that may be implemented to control such threats. Examples of common hazards include microorganisms naturally found in meat or poultry products (i.e. *Campylobacter*, *Salmonella*), chemicals that are unintentially added to food (i.e. pesticides, cleaners), or foreign materials that are accidentally found in food (i.e. metal, plastic).

PRINCIPLE #2: Identify the critical control points (CCPs)

A critical control point (CCP) refers to a point, step, or procedure in the food process during which control measures may be applied to prevent, eliminate, or reduce hazards. An example of a CCP is the procedure of cooking poultry to 165° F to destroy microorganisms that may be present.

PRINCIPLE #3: Establish critical limits for each CCP

Critical limits are defined as the maximum or minimum value at which a biological, chemical, or physical hazard must be controlled at a given CCP to ensure food safety. An example of a critical limit includes holding temperatures, such as the minimum hot holding temperature of 140° F or the maximum cold holding temperature of 41° F.

[†] FDA. HACCP: A State-of-the-Art Approach to Food Safety. http://www.cfsan.fda.gov/~lrd/bghaccp.html

PRINCIPLE #4: Establish monitoring procedures

Monitoring procedures are those procedures that check, measure, and document the food process at a given CCP. An example of a monitoring procedure is the routine observation and recording of cooking times and temperatures.

PRINCIPLE #5: Establish corrective actions

When deviations or problems are identified through monitoring, corrective actions are initiated. An example of a corrective action is the disposing of food if the minimum cooking temperature is not met.

PRINCIPLE #6: Establish recordkeeping procedures

Recordkeeping is essential for the documentation of monitoring procedures, hazards identified, and actions taken to correct potential problems. Moreover, recordkeeping ensures that regulatory requirements are met.

PRINCIPLE #7: Establish verification procedures

Verification procedures are necessary to evaluate a HACCP system and determine if the system is working properly. Verification often involves the testing and reviewing of specific steps, quality control and assurance of equipment and procedures, and annual assessments.

Application of HACCP Principles during an Environmental Investigation

When a foodborne disease outbreak is identified in a food service establishment, food inspectors conduct an inspection that is based on the HACCP principles. Food inspectors follow the food process in the establishment, paying close attention to the preparation of suspect foods or foods implicated in the foodborne disease outbreak.

The following paragraphs describe the general procedures of a HACCP inspection during a foodborne disease outbreak investigation.

Introduction and purpose

Upon arrival at the food service establishment, the inspector should introduce himself to the person in charge and explain the purpose of the inspection.

Identification of ingredients and steps

The inspector should review the menu and identify the ingredients and steps involved in the receiving, storage, preparation and service of suspect food(s). The inspector should obtain recipes for all suspect food(s), identify the ingredients, and collect information about the source. The inspector should also pay close attention to potentially hazardous foods and high-risk preparation factors.

Identify critical control points

Based on the observations made, the inspector should identify critical control points and corrective actions to reduce potential hazards. Microbiological hazards account for the majority of foodborne illness; therefore, emphasis should be placed on contamination, survival, and growth/toxin production risks at these points.

Observe suspect food(s) through establishment

The inspector should observe the suspect food(s) and record the procedures conducted through the operation — from receipt of food from the delivery truck to consumption by the consumer. All risk factors should be observed, including the food source, cooking and holding procedures, potential contamination factors, and poor personal hygiene. Inspectors should have the proper equipment (e.g., thermometers) to assist with these observations. Written documentation on how food(s) were handled and what equipment was used should be completed. Observation and documentation of who handled the food(s) during each preparation step should also be done to help determine if a specific food handler or particular role may have contributed to illness. A flow chart should be developed as a visual tool of the process.

Monitoring and corrective action procedures

Monitoring procedures and corrective actions should be established. These should be discussed in a brief exit interview with the person in charge.

Submit paperwork

Inspectors should write and submit a HACCP inspection report, complete with flow charts, recommendations, and other appropriate paperwork to their supervisor and the epidemiology investigator. Following submission of the report, the inspector should return to the food service establishment to present the report and discuss recommendations with the person in charge.

APPENDIX K

Fact Sheets

- **K.1** Campylobacteriosis
- K.2 E. coli O157:H7
- **K.3** Hepatitis A (Infectious Hepatitis)
- K.4 Hepatitis A (Infectious Hepatitis) and Foodhandlers
- **K.5** Listeriosis
- **K.6** Norwalk-like Viruses
- **K.7** Noroviruses and Foodhandlers
- **K.8** Salmonellosis
- **K.9** Shigellosis



Campylobacteriosis

Health Education Facts

How Common is Campylobacteriosis?

Campylobacter is the most common bacterial cause of diarrheal illness in the United States. Many more cases go undiagnosed or unreported, and campylobacteriosis is estimated to affect over 2 million persons every year. Most cases of campylobacteriosis are associated with handling raw poultry or eating raw or undercooked poultry meat. It can also be transmitted from contaminated water or unpasteurized milk. A very small number of Campylobacter organisms (fewer than 500) can cause illness in humans. Even one drop of juice from raw chicken meat can infect a person. The organism is not usually spread from person to person.

What are the Symptoms?

Most people who become ill with campylobacteriosis get diarrhea, cramping, abdominal pain, and fever within 2 to 5 days after exposure to the organism. The diarrhea may be bloody and can be accompanied by nausea and vomiting. The illness typically lasts 1 week. Some persons who are infected with *Campylobacter* don't have any symptoms at all. In persons with compromised immune systems, *Campylobacter* occasionally spreads to the bloodstream and causes a serious life-threatening infection.

The majority of individuals who get campylobacteriosis recover completely within 2 to 5 days, although sometimes recovery can take up to 10 days, although prolonged illnesses and relapses may occur in adults.

Some Tips for Preventing Campylobacteriosis

- Cook all poultry products thoroughly. Make sure that the meat is cooked throughout (no longer pink), any juices run clear, and the inside is cooked to 165°F
- If you are served undercooked poultry in a restaurant, send it back for further cooking
- Wash hands with soap before, and after, handling raw foods of animal origin
- Use separate cutting boards for foods of animal origin and other foods
- Carefully clean all cutting boards, counter-tops and utensils with soap and hot water after preparing raw food of animal origin
- Avoid consuming unpasteurized milk and untreated surface water
- Make sure that persons with diarrhea, especially children, wash their hands carefully and frequently with soap to reduce the risk of spreading the infection
- Wash hands with soap after having contact with pet feces

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handout #30

There are hundreds of strains of the bacteria *Escherichia* coli. These strains are commonly found in the intestines of healthy persons and animals. A particular strain, E. coli 0157:H7, also known as 0157, can cause severe illness and even death. First identified as a cause of human illness in 1982, this bacterium is increasingly being detected throughout the developed world.

What are the symptoms?

Persons infected with this type of E. coli can develop severe diarrhea and painful abdominal cramps. The diarrhea is often bloody. For most the illness subsides in five to ten days. However, for some, the infection can lead to a condition known as hemolytic uremic syndrome, WS) in which the kidneys fail, and other complications including seizure and stroke can occur.

E. coli 0157 is the principal cause of HUS, and HUS is the primary cause of acute kidney failure in children. Less than ten percent of the infections lead to HUS, but persons with this illness often require intensive care, blood transfusions and kidney dialysis to survive. Most do survive this condition, but some may have high blood pressure and kidney problems later in life.

Where does E. Coli 0157:H7 come from?

Major sources are undercooked ground beef and raw milk. The 0 157 bacterium is present in the intestines of cattle, which during slaughter may come into contact with the ground meat product. The bacteria are killed when meat is thoroughly cooked, but can survive in meat that is rare or inadequately cooked. The lesson for prevention is to cook ground beef to an internal temperature of at least 155°F.

Bacteria present in the cow's udders or in milking equipment can be passed into raw milk, but pasteurization kills the bacteria.

How else can the bacteria be passed on?

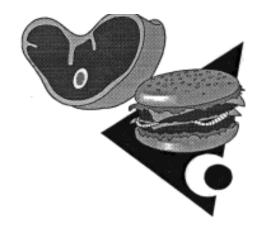
Since waterborne transmission of 0157 has been documented, only chlorinated or carbonated water should be considered safe. Also, the organism is easily passed from person-to-person, and for this reason is a concern in daycare settings. Frequent hand washing with soap will prevent transmission.

Who is at the greatest risk?

Those people primarily at risk of severe consequences of infection are children under five years of age and the elderly.

How widespread is the bacterium?

Preliminary estimates indicate that as many as 20,000 cases of infection from E. coli 0157:H7 occur in the United States each year. The infection is common in Canada, and is increasingly reported in Europe and Japan.



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Handout #34 Revised 01/03



Hepatitis A (Infectious Hepatitis)

Health Education Facts

What is hepatitis A?

Hepatitis A is a virus transmitted by the fecal-oral route. This means that you must get something in your mouth which is contaminated with stool from an infected person.

Who gets hepatitis A?

Most infections result from contact with a household member or sexual partner who has hepatitis A.

Sometimes infection results from eating food or drink which is contaminated with the hepatitis A virus. Once a person recovers from hepatitis A, the person is immune for life and no longer contagious.

How soon after exposure do hepatitis A symptoms appear?

On average, symptoms appear one month after exposure and may include vomiting, diarrhea, and jaundice (whites of the eyes and skin become yellowish). The contagious period lasts from two weeks before to one week after the jaundice starts.

When symptoms are experienced it is important to seek medical care. Since there are several types of hepatitis, a blood test is needed to determine which type is present. Infected children should stay home from school and day care for 10 days following the onset of illness.

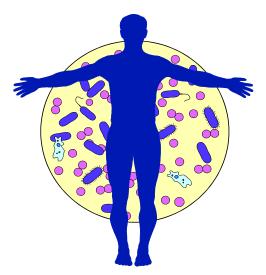
Diarrhea and vomiting can be caused by other things than hepatitis. Adults with moderate to severe gastrointestinal symptoms, particularly diarrhea lasting more than four days, should have a stool culture done through a physician or local health department.

What is the treatment for hepatitis A?

Rest and a balanced diet are usually all that is needed. There is no medication to treat hepatitis A.

How can hepatitis A be prevented?

Handwashing with soap after toileting and diapering is one effective way to prevent the spread of hepatitis A. Gamma globulin (IG) can help prevent infection, and is recommended for people who live in the same house as a person with hepatitis A, to sexual contacts of a person with hepatitis A, or to other children in the same day care center with a child with hepatitis A. IG is NOT given to casual contacts of a person with hepatitis A, such as friends or coworkers, because the risk of infection in these situations is extremely small.



Hepatitis A vaccine is recommended for travelers to countries where hepatitis A is a common infection, and for high-risk adults in this country. Hepatitis A vaccine protects the person who receives it after about one month from the date it is administered. It is not useful for people who have already been exposed to the virus.

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Handout #33 Revised 01/03



Hepatitis A (Infectious Hepatitis) and Food Handlers

Health Education Facts

What is hepatitis A?

Hepatitis A is a virus that causes liver disease. The disease is mild in children, but can be more severe in adults and can cause infected individuals to miss up to 6 weeks of work.

What are the symptoms of hepatitis A?

Symptoms often appear about one month after infection, but can develop anytime between 2 to 6 weeks after infection. Infected persons may experience fever, extreme weakness, loss of appetite, nausea, vomiting, stomach cramps, and diarrhea. These symptoms are usually followed by a few days of dark, "tea-colored" urine and jaundice (yellow skin and eyes).

How is hepatitis A spread?

The hepatitis A virus is found in the feces (stool) of infected persons. Hepatitis A virus is not spread through blood, but is spread primarily through the fecal-oral route. Infected food handlers may carry the virus on their hands and may contaminate readyto-eat food when they do not use good handwashing practices after every restroom visit and have bare hand contact with readyto-eat food. For the disease to spread, the virus must enter the mouth of a person who has not had hepatitis A before or is not immunized against hepatitis A. Good handwashing after toileting and no bare hand contact with ready-to-eat food can greatly reduce the spread of hepatitis A from an infected food handler to others.

How long can an infected person spread hepatitis A?

An infected person can spread the virus for 1 to 2 weeks before symptoms start and up to 10 days after jaundice. Therefore, an infected person may potentially shed hepatitis A virus for almost one month.

What is the treatment for hepatitis A?

There is no specific treatment for hepatitis A. Bed rest and avoiding alcohol, drugs, and over-the-counter medicines is recommended for a faster recovery. Once recovered, a person cannot spread the disease further, is immune for life and cannot be infected with hepatitis A again.



Are food handlers at higher risk for hepatitis A?

Food handlers are not at higher risk than other persons for becoming infected with hepatitis A. However, infected food handlers are at higher *public health risk* for spreading hepatitis A to others. Food handlers have the potential to infect hundreds of people if they work while infected with hepatitis A, do not use good handwashing techniques, and have bare hand contact with ready-to-eat food.

What should be done if a food handler is experiencing symptoms of hepatitis A?

If a food handler is experiencing symptoms of hepatitis A, the food handler should be excluded from food handling immediately, should seek medical care, and should not return to food handling until 2 weeks after the beginning of the illness.

What should be done for co-workers of a food handler infected with hepatitis A?

Co-workers, who worked the same days and shifts of a food handler infected with hepatitis A, should not be allowed to handle food until they receive a shot of immune globulin (IG) to help prevent hepatitis A **or** show proof of previous infection or previous vaccination. The co-workers may return to food handling if they receive IG or provide proof of immunity and are not experiencing any symptoms. If the co-workers refuse IG, they are not allowed to handle food for 50 days starting from their last contact to the infected food handler during the time when the food handler was still contagious.

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Hepatitis A and Food Handlers (continued)

Health Education Facts

If a person receives Immune globulin (IG), is this person immune for life?

No. IG protects a person who has not been infected with hepatitis A for up to 3 months. IG provides protection if it is given before exposure to the hepatitis A virus **or** if it is given within 14 days of exposure to the virus. However, getting IG does not guarantee complete protection against hepatitis A, especially if a person has been exposed to the virus multiple times. IG may make the symptoms milder in a person already infected with hepatitis A.

Are there any health regulations for food handlers infected with hepatitis A?

Yes. To protect the public's health, food handlers infected with hepatitis A cannot handle food until 2 weeks after they started having symptoms. (K.A.R. 28-1-6.) All food handlers should also follow the Kansas Food Code 3-301.11, which prohibits bare hand contact with ready-to-eat food by any food worker.

What are the potential consequences of a food handler who works while infected with hepatitis A?

A food handler who works while infected with hepatitis A has the potential to infect hundreds of customers who have eaten food prepared by the infected worker. The risk of spreading the hepatitis A virus and other diseases increases if the infected food handler has bare hand contact with ready-toeat food. This risk increases even more when an infected food handler has bare hand contact with ready-to-eat food and does not wash his/her hands properly after toileting. When customers have eaten food prepared by a worker infected with hepatitis A, the media is often the most efficient way of informing customers of their possible risk of and treatment options for infection with hepatitis A. This customer recall may affect a restaurant's business, reputation, and financial stability.

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How can food handlers prevent the spread of hepatitis A?

- Food handlers should never work while ill with fever, nausea, vomiting, stomach cramps, or diarrhea. If a food handler is experiencing any of these symptoms, he/she should immediately stop working, inform the manager, and seek medical care.
- 2. Food handlers should always use good handwashing techniques, especially after using the restroom. The proper handwashing method is the following:
 - (a) Use soap and warm, running water;
 - (b) Rub hands together vigorously for 20 seconds;
 - (c) Wash all surfaces, including backs of hands, wrists, between fingers, tips of fingers, under fingernails;
 - (d) Rinse hands well; and
 - (e) Dry hands with a paper towel, if possible.
- 3. Food handlers should not have bare hand contact with ready-to-eat food. If gloves are used, hands must be washed before putting on gloves. Gloves should be used for only one task and then be discarded. Gloves should also be replaced with clean gloves whenever food preparation has been interrupted.

NOTE: Glove usage does not replace the need for food handwashing practices.

4. A vaccine that protects against hepatitis A virus is available and can be used to prevent infection in food handlers.

Is there a vaccine that protects against hepatitis A?

Yes. The hepatitis A vaccine was licensed in the U.S. in June 1995 and is safe and effective for the prevention of hepatitis A. The vaccine is a two-dose series, with the second dose given 6 to 12 months after the first dose. Nearly everyone is protected one month after receiving the first dose, but a second vaccine is needed for lifelong protection.



Listeriosis

Health Education Facts

Listeriosis is a food-borne illness caused by the bacteria Listeria monocytogenes. Although it may cause only a few or no symptoms in healthy people, it can cause serious illness in people with immune system problems, the elderly and pregnant women.

How is Listeriosis Spread?

People get listeriosis by eating food contaminated with the bacteria *L. monocytogenes*. Some types of soft cheeses, undercooked poultry, hot dogs not thoroughly reheated, foods from deli counters, and other ready-to-eat foods are responsible for most reported cases. There have been a few cases of farm workers and veterinarians who have gotten it from animals, but this is not common.

What are the Symptoms?

Healthy people with listeriosis may not have noticeable symptoms. For others, symptoms include fever, fatigue, nausea, vomiting, and diarrhea. Listeriosis, if left untreated, can lead to meningitis (brain infection) and bacteria in the bloodstream. Pregnant women may develop flulike symptoms with complications resulting in miscarriage, stillbirth, or meningitis in their newborn baby. In older children and adults, listeriosis may attack the central nervous system and bloodstream, causing pneumonia and inflammation of the lining of the heart and valves. Abscesses or skin lesions may also appear. Although flu-like symptoms may occur 12 hours after eating contaminated food, it usually takes from one to six weeks for a serious case of listeriosis to develop. The time it takes for symptoms to appear depends on the person's age and general health, the strain of L. monocytogenes, and how much bacteria was eaten.

How Can Listeriosis be Prevented?

Although most people are at very low risk for listeriosis, the risk of this and other food-borne illnesses can be reduced by following the tips listed below:

- Avoid using or drinking unpasteurized milk.
- Keep raw and cooked foods separate when shopping, preparing, cooking, and storing

foods. Bacteria in juices from raw meat, poultry, and fish can contaminate cooked food.



- Wash your hands, knives, and cutting boards after handling uncooked foods.
- Wash raw vegetables well before eating them.
- Thoroughly cook meats, poultry, eggs, and fish.
- Read and follow directions on foods to keep them properly refrigerated. Do not use foods after their expiration date.
- Keep hot foods hot and cold foods cold. Do not keep them unrefrigerated longer than two hours.
- Place leftovers in shallow, covered containers and refrigerated them immediately.
- Keep your refrigerator clean, and keep the temperature in it between 34 and 40 F.
- Pregnant women, the elderly, and people who have weakened immune systems can reduce their risk by following the tips below:
- **F**
- Avoid soft cheeses such as Mexican style (soft, white ethnic cheeses), feta, Brie, Camembert, and blue cheese. There is no need to avoid hard cheeses, processed slices, cottage cheese, or yogurt.
- Reheat leftover foods and ready-to-eat foods such as hot dogs thoroughly until they are steaming hot.
- Although the risk of listeriosis from deli counter food is relatively low, avoid these foods and thoroughly reheat cold cuts before eating them.

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HOW COMMON ARE NORWALK-LIKE VIRUSES?

Only the common cold is reported more frequently than viral gastroenteritis as a cause of illness in the United States. Although viral gastroenteritis is caused by a number of viruses, it is estimated that Norwalk-like viruses are responsible for about 1/3 of the cases in those over 2 years of age. Norwalk-like viruses are increasingly being recognized as a leading cause of foodborne disease. Though first discovered in 1972, it wasn't until the viruses were cloned in 1990 that researchers were able to study them.

HOW ARE NORWALK-LIKE VIRUSES TRANSMITTED?

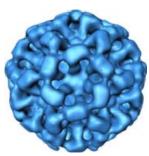
Norwalk-like gastroenteritis is usually transmitted via hands inadequately washed after toileting. The virus is then transferred to food or water. Secondary person-to-person transmission has been documented. Water is the most common source of outbreaks and may include water from municipal water supplies, wells, recreational lakes, swimming pools and water stored aboard ships. Shellfish and salad ingredients are the foods most often implicated in Norwalk-like outbreaks. Ingestion of raw or insufficiently steamed clams and oysters poses a high risk for infection. Foods other than shellfish are contaminated by ill food handlers.

WHAT ARE THE SYMPTOMS?

A mild and brief illness usually develops 24-48 hours after contaminated food or water is consumed and lasts for 24-60 hours. The disease is self-limiting, mild and characterized by nausea, vomiting, diarrhea and abdominal pain. Headache and low-grade fever may occur. Severe illness or hospitalization is very rare. Person-to-person transmission can occur up to 7 days after an affected person has stopped having symptoms.

SOME TIPS FOR PREVENTING NORWALK-LIKE VIRUS INFECTION

- Wash hands with soap and warm water after toilet visits and before preparing or eating food
- Cook all shellfish thoroughly before eating
- Wash raw vegetables before eating
- Food handlers with symptoms of Norwalk-like viruses should not prepare or touch food



Norwalk Virus Electron Micrograph

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Noroviruses and Food Handlers

What are noroviruses?

Noroviruses are members of a group of viruses called caliciviruses also known previously as "Norwalk-like viruses." Infection with norovirus affects the stomach and intestines, causing an illness called gastroenteritis, or "stomach flu." This "stomach flu" is *not* related to the flu (or influenza), which is a respiratory illness caused by influenza virus. In addition, noroviruses are not related to bacteria and parasites that can cause gastrointestinal illnesses. Norovirus is not a "new" virus, but interest in it is growing as more is learned about how frequently noroviruses cause illness in people (see – "Why is norovirus infection important for food handlers?").

What are the symptoms of infection with norovirus?

Norovirus infection causes gastroenteritis, which is an inflammation of the stomach and the small and large intestines. The symptoms of gastroenteritis are nausea, vomiting, and/or diarrhea accompanied by abdominal cramps. Some people also complain of headache, fever/chills, and muscle aches. Symptoms are usually brief and last only 1 or 2 days. However, during that brief period, people can feel very ill and vomit, often violently and without warning, many times a day. Symptoms usually begin 24 to 48 hours after ingestion of the virus, but can appear as early as 12 hours after exposure (see – "How is norovirus spread?"). There is no evidence that sick persons can become long-term carriers of the virus, but the virus can be in the stool and vomit of infected persons, from the day they start to feel ill to as long as 2 weeks after they feel better.

Other infectious and non-infectious agents can cause symptoms similar to those of norovirus gastroenteritis; people who have these symptoms and have questions about the cause of their illness should consult a physician.

How serious is norovirus gastroenteritis?

Norovirus gastroenteritis is usually not a serious illness, and other than drinking liquids to prevent dehydration, there is no specific treatment. Most people recover completely within 1 to 2 days, with no long-term complications of norovirus illness. However, persons who are unable to drink enough liquids to replace those lost with vomiting and/or diarrhea may become dehydrated and require special medical attention. These people include young children, the elderly, and persons of any age unable to care for themselves.

How is norovirus spread?

Noroviruses are found in the stool or vomit of infected people. People can become infected with the virus in several ways, including:

- eating food (see food handler fact sheet) or drinking liquids that are contaminated with norovirus:
- touching surfaces or objects contaminated with norovirus, and then placing their hand in their mouth;
- having direct contact with another person who is infected and showing symptoms (for example, when caring for someone with illness, or sharing foods or eating utensils with someone who is ill).

Food and drinks can very easily become contaminated with norovirus because the virus is so small and because it probably takes fewer than 100 norovirus particles to make a person sick. Food can be contaminated either by direct contact with contaminated hands or work surfaces that are contaminated with stool or vomit, or by tiny droplets from nearby vomit that can travel through air to land on food. Although the virus cannot multiply outside of human bodies, once on food or in water, it can cause illness.

Some foods can be contaminated with norovirus *before* being delivered to a restaurant or store. Several outbreaks have been caused by the consumption of oysters harvested from contaminated waters. Other produce such as salads and frozen fruit may also be contaminated at source.

Why is norovirus infection important for food handlers?

People working with food who are sick with norovirus gastroenteritis are a particular risk to others, because they handle the food and drink many other people will consume. Since the virus is so small, a sick food handler can easily — without meaning to — contaminate the food he or she is handling. Many of those eating the contaminated food may become ill, causing an outbreak.

Outbreaks of norovirus gastroenteritis have taken place in restaurants, cruise ships, nursing homes, hospitals, schools, banquet halls, summer camps, and family dinners — in other words, places where often people have consumed water and/or food prepared or handled by others. It is estimated that as many as half of all food-related outbreaks of illness may be caused by norovirus. In many of these cases, sick food handlers were thought to be implicated.

What can I do to prevent norovirus gastroenteritis?

Many local and state health departments require that food handlers and preparers with gastroenteritis *not* work until 2 or 3 days after they feel better. In addition, because the virus continues to be present in the stool for as long as 2 to 3 weeks after the person feels better, strict hand washing after using the bathroom and before handling food items is important in preventing the spread of this virus. Food handlers who were recently sick can be given different duties in the restaurant so that they do not have to handle food (for example, working the cash register or hostessing).

People who are sick with norovirus illness can often vomit violently, without warning, and the vomit is infectious; therefore, any surfaces near the vomit should be promptly cleaned and disinfected with bleach solution and then rinsed. Furthermore, food items that may have become contaminated with norovirus should be thrown out. Linens (including clothes, towels, tablecloths, napkins) soiled to any extent with vomit or stool should be promptly washed at high temperature. Oysters should be obtained from reputable sources and appropriate documentation kept. Washing raw vegetables thoroughly before eating and appropriate disposal of sewage and soiled diapers also help to reduce the spread of norovirus and prevent illness. In small home-based catering businesses or family owned or operated restaurants, sick children and infants in diapers should be excluded from food preparation areas.

How is norovirus gastroenteritis diagnosed?

In special cases, when there is an outbreak of gastroenteritis there is a need to identify norovirus as the cause of the illness. In these cases, norovirus can often be found in stool samples of infected persons by using special tests. Sometimes blood tests looking for antibodies against norovirus are also performed, when the stool tests are inconclusive or were not done. Food handlers will often be asked for a stool sample or even a blood sample to help investigate the cause of an outbreak.

Can a person have norovirus gastroenteritis more than once?

Yes, a person can be infected with norovirus more than once in their lifetime. This is because there are many different noroviruses, and being infected with one type does not prevent infection from another type later. For this reason, it is difficult to develop a vaccine against norovirus.



Salmonella is a bacterium found in contaminated foods, and grows when foods are improperly handled or prepared. After consuming contaminated foods, serious intestinal problems may result.

Symptoms and treatment

Symptoms include fever, cramps, nausea, and diarrhea but may vary depending on the individual and amounts of contamination. Young children, people with special health conditions, and senior citizens are more likely to experience severe symptoms with complications. A doctor or health care worker should determine if treatment is necessary.

How is salmonella spread?

Salmonella bacteria are passed through the intestines of humans, mammals, and birds. Outbreaks of Salmonella have been traced to processed meats, undercooked poultry or poultry products, raw or lightly cooked foods containing egg or egg products (such as homemade ice cream), raw sausages, and unpasteurized milk and dairy products (including dried milk). Foods contaminated with rodent feces, or prepared with improperly cleaned utensils or work space, spread Salmonella infection. People with symptomatic Salmonella infection should not be handling foods.

Prevention Careful fo

Careful food handling is necessary in controlling Salmonella infection. The bacteria grow and survive at room temperature, so proper food temperatures should be maintained. Salmonella is not killed by freezing, but will not grow rapidly below 40°F. Thawing meats and other foods in the refrigerator or cooking them while they are still frozen will lessen the chance of Salmonella infection. Foods must be thoroughly cooked to kill the bacteria.

Utensils and surfaces should be carefully

eating. Hands should be washed immediately after handling diapers.

To avoid food borne illness:

hands after using the restroom and before

washed and sanitized after uncooked foods are

use the same utensils for raw foods and cooked

foods without washing and sanitizing. Careful

hand washing before preparing and handling food is important. Everyone should wash their

handled. Avoid cross contamination -- do not

- Wash hands often, and keep your nails trimmed and clean.
- Keep work areas clean and sanitary.
- Do not use the same utensils for raw and cooked foods without thorough washing and sanitizing.
- Thaw foods in the refrigerator, under running cold water, or as part of the cooking process.
- Keep hot foods hot and cold foods cold.
- Never let foods sit at room temperature for more than two hours.
- Never consume raw eggs or raw egg products.
- Cook poultry thoroughly.

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What causes shigellosis?

Shigellosis is caused by bacteria belonging to the *Shigella*. species. These are: *Shigella dysentefiae 1, Shigella sonnei, Shigella flexneri, and Shigella boydii.*

Who gets shigellosis?

Shigellosis affects people of all ages; however, infants and the elderly are at greater risk of disease.

How is shigellosis transmitted?

Only humans carry *Shigella*. Transmission occurs by the fecal-oral route. The usual mode of transmission is through contaminated hands that transfer the bacteria to food or water. Person-to-person transmission also may occur. Flies may transmit the disease by carrying the bacteria on their legs to food. Dogs that eat human feces may transmit the disease to people, especially children.

What are the symptoms and how soon do they appear?

The incubation period ranges from 12 to 96 hours, but may be as long as one week. Symptoms usually include bloody diarrhea accompanied by fever, nausea, vomiting, abdominal cramps, and painful, involuntary contractions of the anus. Severe cases can result in death. Mild and asymptomatic cases may occur. Illness is often self-limiting lasting four to seven days, occasionally up to weeks or months.

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How is shigellosis diagnosed?

Shigellosis is diagnosed by culturing the stool for the bacterium. This is done through a laboratory test.

How is shigellosis treated?

Antibiotics are used to treat shigellosis and are effective in shortening the course of illness.

What can be done to prevent shigellosis?

Infection and transmission of *Shigella* can be prevented by:

- Consuming water from a safe source.
- Always washing hands after toileting, before preparing or serving meals, and before eating. Wash hands with soap for 20 seconds then rinse with warm running water.
- Washing fruits and vegetables before eating.
- Always washing hands after petting animals and changing diapers.

